# Phylogenetic relationships of Abildgaardieae (Cyperaceae) inferred from chloroplast and nuclear DNA sequences and pollen data

# **Kioumars Ghamkhar**

M.Sc., The University of Isfahan B.Sc., The University of Isfahan

This thesis is submitted for the degree of Doctor of Philosophy of the University of New England

January 2004

The University of New England Armidale, NSW 2351, Australia

To:

**MOJGAN** 

MARAL

&

NAHAL

## Declaration

I certify that the substance of this thesis has not already been submitted for any degree and is not currently being submitted for any other degree or qualification.

I certify that any help received in preparing this thesis, and all sources used, have been acknowledged in this thesis.

Kioumars Ghamkhar

## Acknowledgments

There is no question in my mind that my gratitude for achieving this milestone should begin with my parents. Their tireless provision of the two foods (physical and mental) has been the foundation stone of my success. I have had parents of whom I am so proud. So much of who I am, I owe to them. In particular, thanks to my mum for her ongoing support.

I am deeply grateful to my supervisors Associate Professor Jeremy Bruhl, Adjunct Associate Professor Karen Wilson, and Dr Adam Marchant for being ideal supervisors. I am sincerely thankful for their wise counsel, invaluable advice, ongoing guidance, constructive criticism, encouragement, and patience throughout this study. I was lucky that they were not only my supervisors but also my best friends in Australia.

I extend my profound thanks to the Ministry of Science, Research, and Technology, the Government of Islamic Republic of Iran, for providing a scholarship during the period of my studies. I give thanks especially to Professor Javad Farhoodi for his enormous support.

I extend my sincere thanks to Professors Chris Quinn and John Thomson of Royal Botanic Gardens, Sydney, and Dr Helene Martin of the University of New South Wales for their kind advice during this study.

My sincere thanks are due to Ms Kerri Clarke of the University of New England (UNE). Associate Professor Jeremy Bruhl, and Mr Van Klaphake, for providing herbarium and silica gel plant materials, Adjunct Associate Professor Karen Wilson of Royal Botanic Gardens (RBG), Sydney, and Mr. George Orel of the University of Western Sydney for providing CTAB preserved plant material.

I have experienced a very positive working environment. Thanks to all my colleagues and officemates namely Dr Adam Marchant, Dr Andrew Perkins, Carolyn Porter, Nick Yee, and George Orel for their contributions. My special thanks go to

Dr Adam Marchant and Dr Andrew Perkins, who have been always the first point of help when I was in need. I would, also, like to thank my very special friend, Carolyn Porter, for helping me over the duration of my PhD. Her role in the earliest technical supervisions both in the molecular and SEM work is greatly acknowledged.

I appreciate supports from people at the Plant Sciences section at RBG, and Botany at UNE. My special thanks go to Dr Tim Entwisle, Dr Barry Conn, and Mr Anthony Martin at RBG and Mr Chris Cooper and Mr Douglas Clark at UNE.

I thank the molecular lab meeting group at RBG, Sydney, for the useful meetings and fruitful discussions and specifically Dr Peter Weston for his always useful comments.

I am grateful to my parents-in-law for their invaluable support and encouragement and for always being there for me. In addition, my brother and sisters and entire extended family are thanked for their support.

I have found that my very young and lovely daughter, Maral Ghamkhar, has been a great source of encouragement and motivation. I thank her and wish her the best for being reasonable and patient over the last four years.

Finally, I thank my lovely wife, Mojgan. Her incredible personal sacrifice and support are what has genuinely made all this possible. Without her kind and generous support, in particular, emotionally, I believe that this thesis would never have come into being. My gratitude to her is too deep for words to convey.

I hope that I will honour those individuals who have contributed to my success by carrying the torch high and passing it on with a light that burns brighter than when I received it.

This work received support from Botany, University of New England (N.C.W. Beadle Fund), the Plant Sciences section RBG, Sydney, and the Linnean Society of New South Wales (Joyce W. Vickery Scientific Research Fund).

Kioumars Ghamkhar

## Prologue

#### **Format**

The format of this thesis follows wherever possible the format of *Australian Systematic Botany* except that:

- figures and plates are located throughout the text
- spaces have been placed between paragraphs
- section headings are numbered

#### Thesis structure

Chapter one provides an introduction to the study, reviews previous approaches to taxonomic issues within and among Abildgaardieae and its allies, and outlines the relationships and taxonomic history of the study group. Aims and hypotheses to be tested are stated along with how and why these changed over the course of the study.

Chapter two reviews the process of selection of molecular methods. Theoretical considerations and selection of data sources are reviewed. Various molecular techniques and methods are described and compared and their phylogenetic value discussed.

The use and methodology of *trn*L–*trn*F non-coding region of chloroplast DNA is reviewed in chapter three. The methods and results of phylogenetic analyses for the study group taxa are described and discussed.

Chapter four deals with the use of the ITS region of the nuclear ribosomal DNA and phylogenetic analyses of separate ITS and combined ITS/trnL-trnF data. The results are discussed and compared with the previous studies.

Chapter five provides an overview and history of pollen morphology studies and presents and discusses characters of pollen morphology in the study group and phylogenetic analysis of such data. This chapter also deals with combined molecular (ITS/trnL-trnF) and morphological data using different approaches and analytical methods, and compares the results of separate analysis and combined molecular analysis. Comparison with previous studies is the final part of this chapter.

The last chapter, six, provides an overview of the phylogenetic relationships, implications and limitations of the present study. Future directions and priorities for systematic studies of Abildgaardieae and Arthrostylideae are presented.

# Data availability

The datasets are available from the author on request.

#### **Abbreviations**

AFLP amplified fragment length polymorphism

AMB ambitus

Bp base pair(s)

CI consistency index

cpDNA chloroplast DNA

CTAB hexadecyltrimethylammonium bromide

(cetyl trimethylammonium bromide)

DELTA description language for taxonomy

DNA deoxyribonucleic acid

dNTP(s) deoxyribonucleotide triphosphate(s)

EDTA ethylenediamine tetra-acetic acid

g gram(s)

GBSSI granule-bound starch synt iase

GHz gigahertz

IGS intergenic spacer

ILD incongruence length difference

IR inverted repeat

ITS internal transcribed spacer

LM light microscope/y

LSC large single copy region

min minute(s)

ML maximum likelihood

ml millilitre(s)

mM millimolar

MP maximum parsimony

NJ neighbour-joining

NSW New South Wales

NT Northern Territory

NTS non-transcribed spacer

PAUP phylogenetic analysis using parsimony

PCR polymerase chain reaction

pers. comm. personal communication

pH log of H+ concentration

PHT partition homogeneity test

QLD Queensland

RAPD random amplified polymorphic DNA

RFLP restriction fragment length polymorphism

RI rescaled consistency index.

RNA ribonucleic acid

rpm revolutions per minute

SAW successive approximation weighting

sec second(s)

SEM scanning electron microscope/y

SSM slipped-strand mispairing

SSRs simple sequence repeats

SSC small single copy region

TAPS tris-(hydroxymethyl)-methyl-amino-propanesulfonic acid sodium salt

Taq thermus aquaticus

TBE tris-borate/EDTA electrophoresis buffer

TBR tree bisection reconnection

TEM transmission electron microscope/y

Tris tris (hydroxymethyl)-aminomethane

Tris HCl tris (hydroxymethyl)-aminomethane hydrogen chloride

UV ultraviolet

WA Western Australia

μl microliter(s)

μg microgram(s)

°C degrees Celsius

#### Abstract

The need for a systematic study of the tribe Abildgaardieae became apparent when the boundaries between Abildgaardieae and Arthrostylideae in a previous study using morphological and physiological characters were unclear. Within Abildgaardieae, some species belonging to separate genera overlap in morphological and/or embryological characters (viz. Fimbristylis hispidula, Abildgaardia ovata, Bulbostylis pilosa) and their recognition in separate genera has been questioned.

The focus of this study was to estimate phylogeny for Abildgaardieae and evaluate the circumscription of six genera (49 species) within Abildgaardieae and Arthrostylideae. Parsimony and maximum likelihood analyses were conducted separately on three different datasets (ITS (nuclear ribosomal), *trn*L-F (plastid) DNA sequence, and pollen morphological data). The data sets were also combined and analysed using the same techniques.

The results showed that the three data sets produced different phylogenies. Separate data sets did not clearly resolve the relationships within Abildgaardieae or between Abildgaardieae and Arthrostylideae. By combining the three datasets, much higher resolution of the phylogenetic relationships was observed. ITS and *trnL*-F provided different insights into the relationships within Abildgaardieae and between the tribes disagreeing with the position of Arthrostylideae relative to Abildgaardieae and with the monophyly of *Abildgaardieae*. Pollen morphology provided useful taxonomic characters for the Abildgaardieae--Arthrostylideae complex but it did not resolve the relationships further when analysed alone. When combined with ITS and *trnL*-F, pollen morphology further strengthened *trnL*-F results.

One of the genera of Abildgaardieae, *Bulbostylis*, formed a well-supported 'basal' clade distinct from related genera. Genera (two sampled) of Arthrostylideae were nested between *Bulbostylis* and the remaining genera (three sampled) of Abildgaardieae. Within these genera of Abildgaardieae a highly resolved clade composed of *Fimbristylis*, *Crosslandia* and *Abildgaardia vaginata* emerged indicating the need to redefine the limits of these genera. The data do not support recognition of the taxa within this last clade as distinct genera.

Crosslandia, Abildgaardia, and Fimbristylis formed a clade in all analyses. Total

evidence of molecular and pollen data indicate that Arthrostylideae are embedded in Abildgaardieae and *Bulbostylis* is monophyletic. *Fimbristylis* is monophyletic only with the inclusion of *Crosslandia* and *4. vaginata*. The rest of *Abildgaardia* is monophyletic and sister to this expanded *Fimbristylis*. Relationships within *Fimbristylis* largely do not conform to infrageneric classification.

# Table of contents

Dedication	ii
Declaration	iii
Acknowledgments	iv
Prologue	vi
Abbreviations	viii
Abstract	X
Table of contents	xii
Table of figures	xviii
Table of tables	xx
Chapter 1 (General introduction)	
1.1 Introduction to the taxonomy of Cyperaceae	1
1.1.1 Difficulties in Cyperaceae	2
1.1.1.1 Problem of defining Scirpeae and related tribes	3
1.1.2 Abildgaardieae	4
1.1.2.1 Relationships of Abildgaardieae and its closest tribes	5
1.1.3. Arthrostylideae	7
1.1.4. Genera of Abildgaardieae	9
1.2. Value of non-molecular data	12
1.2.1 Suitability of pollen grains as a source of characters	13
1.2.1.1 Pollen grains in Cyperaceae	13
1.2.2. Embryomorphology of Abildgaardieae	16
1.3. Molecular systematic studies of Abildgaardieae	16
1.3.1 Relationships of Abildgaardicae	17
1.3.1.1. Relationships of Abildgaardieae inferred from separate molecular	ar and
morphological data	18
1.3.1.2. Relationships of Ab Idgaardieae inferred from combined data	18
1.4. Aims of this study	21
Chapter 2 (Molecular systematics)	
2.1 Introduction	23
2.1.1 Problems with morphological data	23
2.1.2 Link between phenotype and genotype	23
2.1.3 Phytochemistry as a data source	24

	2.1.4 Molecular sequence data for phylogenetic reconstruction	24
	2.1.5 Proteins.	25
2.2 T	echniques	25
	2.2.1 PCR technique	25
	2.2.2 DNA sequencing	26
	2.2.2.1 Transition, transversion and codons	26
	2.2.2.2 Character weighting	27
	2.2.2.3 Sequence variation	27
	2.2.2.4. Advantages	29
	2.2.3 Analysing DNA	29
	2.2.3.1 Genes and gene trees	30
	2.2.4 Markers and population variation	31
	2.2.4.1 DNA fingerprinting	32
	2.2.4.1.1. RAPDs	32
	2.2.4.1.2. Microsatellites	32
	2.2.5. Restriction site mapping	33
	2.2.5.1. Limitations	33
	2.2.5.2. Restriction site mapping vs. sequence data	34
	2.2.5.3. Structural changes in restriction site mapping	35
	2.2.5.4. Restriction fragment length polymorphism (RFLP)	35
	2.2.5.5. AFLP as a mixture of restriction site and PCR methods	35
2.3 C	haracter evolution	36
2.4 G	Genomes	36
	2.4.1 Genome inheritance	36
	2.4.2 Nature of cpDNA	37
	2.4.2.1 cpDNA for phylogeny	37
	2.4.2.2 <i>rbc</i> L	37
	2.4.2.2.1. rbcL for deep plant phylogeny	38
	2.4.3 mtDNA.	41
2.5 M	Nolecular clock	41
2.6 S	eparate vs. combined analyses	42
2.7 A	dvantages and disadvantages of various methods and gene regions	43
	2.7.1 Chloroplast gene regions other than <i>rhc</i> L	43
	2.7.1.1 <i>ndh</i> F	43
	2.7.1.1.1. Combined data from rbcL and ndhF	44
	2.2.1.1.2 Variation within ndbF	44

2.7.1.2 matK	.5
2.7.1.2.1. Variation within matK	5
2.7.1.2.2. Comparisor of rbcL and matK	7
2.7.1.2.3. Combined data from rbcL and matK	8
2.7.1.3 Non-coding regions of DNA	8
2.7.1.3.1. Introduction	8
2.7.1.3.2. Variation in non-coding regions4	9
2.7.1.3.2.1. Basis of variation of non-coding regions4	9
2.7.1.3.2.1.1. Slipped-Strand Mispairing (SSM)4	9
2.7.1.3.2.1.2. Stem loop secondary structure 5	0
2.7.1.3.2.1.3. Nucleotide substitutions5	0
2.7.1.3.2.1.4. Intramolecular recombination 5	1
2.7.1.3.3. Taxonomic value of non-coding DNA	2
2.7.1.3.4. Alignment of non-coding DNA	3
2.7.1.3.4.1. Homology	3
2.7.1.3.5. Analysisof non-coding DNA	3
2.7.1.3.6. trnL intron and trnL-trnF intergenic spacer	4
2.7.2 Nuclear genes (regions)	5
2.7.2.1 Nuclear ribosomal DNA (nrDNA)	5
2.7.2.1.1. Characteristics	6
2.7.2.1.2. ITS	6
2.7.2.1.3, 5S-NTS	9
2.7.2.1.4. Value of high copy nuclear regions	9
2.7.2.2 Other nuclear genes.	0
2.7.2.2.1. waxy gene	0
2.7.2.2.2. <i>Chs</i>	0
2.7.3 Review of analytical methodology6	0
2.8 Objective of this study6	4
Chapter 3 (Phylogenetic relationships within the Abildgaardieae– Arthrostylideae	
group: trnL intron and trnL-trnF Intergenic spacer)	
3.1 Introduction	
3.2 Materials and methods6	
3.2.1 Plant material 6	
3.2.1.1 Sampling	
3.2.2 DNA extraction, amplification and sequencing	
3.2.2.1 DNA isolation	8

3.2.3 Alignment and gap coding	70
3.2.4 Phylogenetic analyses	71
3.2.5 Measures of character fit and character weighting	77
3.2.5.1 Character weighting	77
3.3 Results	79
3.3.1 Characteristics of the trnL-F intergenic spacer region and the trnL intron.	80
3.3.2 Phylogenetic analysis	82
3.4 Discussion	88
Chapter 4 (Internal transcribed spacers of the nuclear genome (ITS) and combin	ed
ITS/trnL-trnF data for phylogenetic studies in the Abildgaardieae)	
4.1 Introduction	91
4.2 Materials and methods	91
4.2.1 Plant material	91
4.2.2 nrDNA region analysed	91
4.2.3 DNA extraction, amplification and sequencing	92
4.2.3.1 Laboratory procedures	92
4.2.4 Sequence analysis, methods of analysis and testing	93
4.2.4.1 Combined ITS/trnL-I data	95
4.3 Results	97
4.3.1 ITS structure, size, and composition	97
4.3.2 ITS phylogenetic analysis	98
4.3.3. The phylogenetic analysis of the ITS/trnL-F combined dataset	101
4.3.4 Comparison of exchange rates of the ITS region versus trnL-F region and	trnL
intron	107
4.4 Discussion	111
4.4.1 ITS in Abildgaardieae	111
4.4.1.1 Relationships and monophyly	112
4.4.2 Combining data	114
4.4.2.1 Resolution of clades from combined cpDNA and nrDNA data	114
4.4.3 Conclusion	117
Chapter 5 (Micromorphological pollen data join molecular data)	
5.1 Introduction	119
5.1.1 A review of palynological studies	119
5.1.1.1 Early studies (1682–1969)	119
5.1.1.2 More recent studies (1970 to present)	120

	5.1.1.3 Pollen studies in monocotyledons	.121
	5.1.2 Combined datasets	. 122
	5.1.3 Use of pollen data in phylogenetic studies	. 123
	5.1.3.1 Pollen studies in Cype aceae	. 125
	5.1.4 Different approaches in combining data	. 126
	5.1.5 Aspects of pollen morphology	.129
	5.1.5.1 Pollen terminology	.129
	5.1.5.2 The character states	. 130
	5.1.5.2.1. Apertures and their phylogenetically significant aspects	130
	5.1.5.2.2. Apertures and the effect of acetolysis	.131
	5.1.5.2.3. Exine outer ornamentation	131
	5.1.5.2.4. Shape	133
	5.1.5.2.5. Size	133
	5.1.6 Palynological characterisation of taxa above the rank of order	133
	5.1.7 Preparation method for scanning electron microscopy	.134
	5.1.8 Pollen grains in this study	135
5.2 N	Aaterials and methods	135
	5.2.1. Treatment with Agepon	136
	5.2.2 Treatment with boiling water	136
	5.2.3 Fixation with alcohol	137
	5.2.4 Scanning electron microscopy (SEM)	137
	5.2.5 Light microscopy (LM)	137
	5.2.6 Pollen terminology in this study	138
	5.2.6.1 Apertures	138
	5.2.6.1.1. Existence of aperture	139
	5.2.6.1.2. Aperture type	139
	5.2.6.1.3. Aperture number	139
	5.2,6.1.4. Aperture position	140
	5.2.6.1.5. Aperture margin	140
	5.2.6.2 Shape and polarity of grains	140
	5.2.6.2.1. Pollen shape	140
	5.2.6.2.2. Polarity of pollen	141
	5.2.6.3 Sculpturing	141
	5.2.6.3.1. Distinctness of exine sculpturing	141
	5.2.6.3.2. Sculpturing type	141
	5.2.6.3.3. Sculpturing ordination	142
	5.2.6.3.4. Sculptural density	142

5.2.6.4 Size of pollen	142
5.2.6.5 AMB = ambitus (outline)	143
5.2.6.6 Areolae	143
5.2.6.7 Perforation of tectum	143
5.2.6.8 Contact area	143
5.2.6.9 Informative characters	144
5.2.7 Choosing terminal taxa	144
5.2.8 Phylogenetic analyses	144
5.2.8.1 Combined molecular-morphological analysis	145
5.2.9. Producing a pollen key for the study group	148
5.3 Results	148
5.3.1 Pollen morphological data	148
5.3.2. Morphological analyses	153
5.3.3. Combined analyses	163
5.4. Discussion	166
5.4.1. Pollen studies	166
5.4.1.1 Proposed pollen types	171
5.4.2 A key to the pollen of the Abildgaardieae-Arthrostylideae group	171
5.4.3. Combined morphological–melecular studies	175
Chapter 6 (General discussion)	
6.1 Towards a new resolution	178
6.1.1. Subfamilial and suprageneric delimitaion	178
6.1.2. Generic relationships in Abildgaardieae and Arthrostylideae	179
6.1.3. Sections within Fimbristylis.	181
6.1.4. Final message	182
References	183
Appendix 1. Taxa included in the phylogenetic study of Abildgaardieae	240
Appendix 2. Taxon by character data matrix of the pollen morphological study	243
Appendix 3. Data set for phylogenetic analysis of trnL-trnF	246
Appendix 4. Data set for phylogenetic analysis of ITS	277
Appendix 5. DNA isolation method used in this study	291
Appendix 6 DNA purification method (modified by A. Marchant) used in this study	292

# Table of figures

Figure number	Title	Page
Figure 1.1	Pollen grains of Lepironia mustonata and Carex pringlei	16
Figure 1.2	Diverging groups of Cyperaceae from Rhynchosporeae headed for Abildgaardica	e 17
Figure 1.3	Part of a tree from combined data using weighted characters in Cyperaceae	20
Figure 2.1	Gene splitting before or after population splitting in polymorphic populations	31
Figure 2.2	Genetic circle map of the Oryza sativa chloroplast genome drawn to scale	39
Figure 2.3	Type 1b gap as described by Golenberg et al. (1993) and Hoot and Douglas (1998)	53
Figure 2.4	Different parts of the non-coding trn1F intergenic spacer and trn1. intron intron	55
Figure 2.5	Variable versus conserved sites in the nrDNA	57
Figure 3.1	The primers used in this study to amplify trnL-F region	76
Figure 3.2	Alignment of the intergenic spacer of the trn1F region for a few sampled taxa	77
Figure 3.3	Simultaneous PCR amplification of chloroplast DNA used for a few taxa	81
Figure 3.4	Strict consensus tree of 1000 most parsimonious trees from the trnL-F region data	a 83
Figure 3.5	Part of the nucleotide arrangements of the trnl. intron, in a few sampled taxa	84
Figure 3.6	Phylogram of one of the most parsimonious trees for sampled taxa	85
Figure 3.7	Strict consensus tree of 280 roost parsimonious trees of trn1F region	86
Figure 3.8	One of the 9 cladograms found by successive approximation weighting	87
Figure 3.9	Maximum likelihood tree using a few taxa of the study group	88
Figure 4.1	Primers used in this study to amplify ITS region	93
Figure 4.2	Strict consensus tree from 24 most parsimonious trees for the ITS data set	103
Figure 4.3	One of the three cladograms found by SAW of ITS data	104
Figure 4.4	$\mathbf{M}\mathbf{L}$ tree of the Abildgaardicae-Arthrostylideae complex using a few taxa.	105
Figure 4.5	Comparisons of the number of practical mutations of the three DNA regions	106
Figure 4.6	Strict consensus tree from twelve most parsimonious trees for the combined data	set 1 () 8
Figure 4.7	Cladogram of the only tree found by SAW of combined data	109
Figure 4.8	NJ distance tree from the combined molecular data set	110
Figure 5.1	Examples of exine ornamentation (topography)	132
Figure 5.2	${f SEM}$ micrograph of ${\it Bulbos:vlis\ barbata}$ showing lateral colpoids and distal ulcus	154
Figure 5.3	SEM micrograph of Bulbos:ylis sp. nov showing distal ulcus	154
Figure 5.4	SEM micrograph of Crosslandia setifolia showing lateral colpoids and distal ulcus	154
Figure 5.5	SEM micrograph of Bolboschoenus culdwellii	154

Figure 5.6	SEM micrograph of Fimbristylis densa	154
Figure 5.7	SEM micrograph of Fimbristy lis punctata	154
Figure 5.8	SEM micrograph of Fimbristy lis rara showing lateral colpoids	154
Figure 5.9	SEM micrograph of Fimbristy lis sphaerocephala showing 3 lateral colpoids	154
Figure 5.10	SEM micrograph of Fimbristylis lanceolata showing ridges	155
Figure 5.11	SEM micrograph of Actinoscnoenus sp. showing distal porus	155
Figure 5.12	${f SEM}$ micrograph of <i>Bulbostyis striatella</i> showing distal ulcus and three colpoids	155
Figure 5.13	SEM micrograph of Schoenoplectus literalis. proximal view	155
Figure 5.14	SEM micrograph of Actinoschoenus sp. without colpoids	155
Figure 5.15	SEM micrograph of Abildgaardia pachyptera with colpoids and no distal ulcus	155
Figure 5.16	SEM micrograph of Fimbristylis cymosa: proximal view	155
Figure 5.17	SEM micrograph of Fimbristylis neilsonii (WA form)	155
Figure 5.18	SEM micrograph of Arthrost; lis aphylla: proximal view	156
Figure 5.19	SEM micrograph of Eleochasis cylindrostachys	156
Figure 5.20	SEM micrograph of Abildgaardia pachyptera with the verrucate sculpturing	156
Figure 5.21	SEM micrograph of Crosslavdia setifolia NT2: verrucate sculpturing	156
Figure 5.22	SEM micrograph of Fimbris:ylia sphaerocephala: verrucate sculpturing	156
Figure 5.23	SEM micrograph of Fimbris: ylis cephalophora: scabrate sculpturing	156
Figure 5.24	SEM micrograph of Bulbosy lis sp. nov.: granulate sculpturing	156
Figure 5.25	SEM micrograph of Bulbost lis sp. nov.: granulate sculpturing	156
Figure 5.26	SEM micrograph of Eleocharis cylindrostachys: sculpturing and perforation	157
Figure 5.27	SEM micrograph of Bolboschoenus caldwelii: distal ulcus	157
Figure 5.28	SEM micrograph of $Bulbostylis barbata$ : perforation, ridges, and aperture	157
Figure 5.29	SEM micrograph of Bulbostylis sp. nov.:colpoids and contact area	157
Figure 5.30	SEM micrograph of Bulbosivlis sp. nov.: four out of six lateral colpoids	157
Figure 5.31	SEM micrograph of Abildguardia schoenoides: very fine lateral colpoids	157
Figure 5.32	$\mathbf{S}\mathtt{EM}$ micrograph of <i>Eleocharis cylindrostachys</i> showing five lateral colpoids	157
Figure 5.33	SEM micrograph of Fimbrictylis microcarya: small pollen and colpoids	157
Figure 5.34.	Strict consensus tree of a parsimony analysis of pollen characters	159
Figure 5.35	Strict consensus tree of pollen characters using a constraint	161
Figure 5.36	Strict consensus of pollen characters using the best constraint tree	162
Figure 5.37	Strict consensus tree for combined molecular and pollen datasets	165

## Table of tables

Table number	Title	Page
Table 1.1	Comparison of the recent suprageneric classifications of Cyperaceae	2-1
Table 1.2	Comparison of the intratribal classifications of Abildgaardicae	6
Table 1.3	Suggested relationships and placement of the genera of Arthrostylideae	9-1
Table 2.1	Taxa, and their sequence length used in Hilu and Liang's (1997) study	46
Table 3.1	Primers used for the amplification of two non-coding regions of cpDNA	69
Table 3.2	TA/AT replicates in the trnL trnF IGS among species of Abildgaardicae	82
Table 4.1	Primers of two ITS regions and the 5.8 S coding region between them	93
Table 4.2	Potentially informative indels for relationships within the ingroup	99
Table 4.3	Comparison of genetic evolution in the plastid and nuclear regions	106
Table 4.4	Sequence characteristics of the trnL intron, trnL-F IGS, and the ITS region	107
Table 4.5	The placement of the species sampled in this study, in sections of Kern (1974)	116
Table 5.1	Palynological characters used in the analyses and their Consistency Indices (CI)	149
Table 5.2	Pollen apomorphics of genera and tribes Arthrostylideae, Fuireneae, and Scirpeae	152
Table 5.3	Summary of a few features for pollen grains of the study group based on CI	154-
Table 5.4	Comparisons of the different data sets	164
Table 6.1	Characters used by Kern for delimiting sections in Fimbristylis s.s. and close gener	a 181-1