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Pesticide mixtures in soil: a global outlook

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E-mail: fiona.tang@sydney.edu.au and federico.maggi@sydney.edu.au**Keywords:** agrochemicals, contamination, pesticide residueSupplementary material for this article is available [online](#)**Abstract**

In modern agriculture, pesticides are used in combinations to protect crops. The co-existence of multiple pesticides in soil can threaten the soil biodiversity that maintains ecosystem services, hence further posing a long-term risk on food security. Here, we introduce an assessment of global soil contamination by the residue of pesticide mixtures in nine cropping systems using a fully mechanistic, spatially explicit, and time-resolved model at $0.5^\circ \times 0.5^\circ$ spatial resolution (approximately 55×55 km at the equator) fed with georeferenced agricultural quantities, soil properties, and hydroclimatic variables. We found that 8.3 million km² of treated land have more than one detectable pesticide, with pendimethalin, glyphosate, paraquat, chlorpyrifos, and chlorothalonil being the five most frequently detected. The highest pesticide mixture content was found in the ‘orchards and grapes’ cropping system (95th percentile at 7.3 mg kg soil⁻¹). Globally, the pesticide mixture in the topsoil of approximately 1.88 million km² exceeded 1 mg kg soil⁻¹ for more than 180 d in a year. We estimate that 0.2 million tonnes of pesticides leach below the root zone each year globally, with glyphosate contributing the greatest fraction. The major hotspots of soil pesticide contamination are located in South America and Asia, mainly in Brazil, Argentina, Chile, China, Malaysia, and Japan. Our study shows that the accumulation of pesticide mixtures in soil is a global environmental issue that has to be explicitly accounted for in the sustainability assessment of agricultural production. We propose the use of mechanistic modelling as a tool to aid in designing pesticide management strategies and minimise residue contamination.

1. Introduction

Intensive agriculture uses a variety of pesticides in high amounts to protect crops and secure food production. As of the year 2017, approximately 4.1 million tonnes of pesticides were used globally, corresponding to an annual per capita consumption of 0.5 kg person⁻¹ yr⁻¹ and a market value of USD 71 billion [1]. These figures include more than 500 active ingredients distributed across multiple chemical and functional classes [2] and spanned over 20 000 commercial pesticide products with different formulations [3].

The intensive use of pesticides exerts substantial stress on soil health by affecting soil biota responsible for maintaining soil functions. Many laboratory studies reported a shift in soil microbial community and a decrease in microbial growth and enzymatic activities as a result of exposure to single pesticides [4–8]. Ammonia-oxidising bacteria and archaea are the most sensitive to pesticides, as studies consistently

observed a significant decline in their abundance in pesticide-contaminated soil samples [6, 9, 10]. Karas *et al* [6] also found that pesticides decrease the abundance of sulphur-oxidising bacteria that convert reduced sulphur compounds to sulphate and cause adverse effects on enzymes responsible for mediating the phosphorus cycle. Both ammonia and sulphur oxidisers are key drivers of nutrient cycles that transform nitrogen and sulphur into forms available for microbial and plant uptake, playing a vital role in sustaining soil fertility. In addition to the negative impacts on soil microflora, pesticides affect several earthworm functions by causing disruption to their enzymatic activity, increasing mortality, reducing fertility, altering their feeding behaviour, and decreasing the overall community biomass [11].

More dreadfully, the simultaneous exposure to multiple pesticides can lead to synergistic adverse effects that can deviate from the additive toxicity of single pesticides; though, our current understanding of the response of soil biota to pesticide mixtures

is still inadequate [11, 12]. For example, Yang *et al* [13] observed a clear synergistic effect on earthworms exposed to a quaternary mixture of chlorpyrifos, fenobucarb, clothianidin, and acetochlor, whereas van Hoesel *et al* [14] found that the effects of insecticides and fungicides on earthworm activity were intensified with the simultaneous presence of herbicides. It thus appears that the inputs intended to protect crop production—the pesticides—are potentially causing threats to food security in the long-term.

Despite the negative impacts observed on soil biota, pesticide contamination in soil remains loosely regulated. A review by Jennings and Li (2014) [15] reported that only 174 jurisdictions within 54 United Nations member states promulgated some regulatory guidance values to specify the maximum allowable pesticide residues in soil. Less than 30% of those jurisdictions regulate for more than 100 pesticides, and the guideline values for the same pesticide can vary from 2 to 10 orders of magnitude [15]. In addition, studies also emphasised that the current environmental risk assessment framework used by the European Food Safety Authority (one of the most stringent regulatory bodies) to authorise pesticide commercialisation is insufficient to protect soil ecosystem services [7, 16].

Owing to the lack of regulation, large-scale quantification and monitoring of pesticide residue in the soil are limited. Most studies quantified only a few pesticide substances and these are confined to national- or field-scales [17–19], with Silva *et al* (2019) [20] being one of the most comprehensive studies that covers 76 pesticide residues across 11 European Union member states. While the lack of regulation discourages investments in monitoring campaigns, a comprehensive overview of the current state of global pesticide soil residue is crucial to convey the urgency of adopting sustainable pesticide use, increase public awareness, and substantiate the need for more stringent regulation.

This study puts forth the first global estimate of pesticide residue in agricultural soil. We target to identify the hotspots for pesticide contamination in the topsoil (TS) and leaching below the root zone (BRZ). To this end, we estimate the soil residues of the 92 most used pesticides in nine cropping systems using a mechanistic dynamic model fed with georeferenced databases that include agricultural practice, soil properties, and hydroclimatic variables. The model is solved at $0.5^\circ \times 0.5^\circ$ spatial resolution (approximately 55×55 km at the equator) along a four-layer vertical soil profile down to BRZ, and spanned over a 48 year time window. This study covers 11.85 million km² of croplands within 155 countries and excludes pastures. We benchmark our model outputs against previously published field measurements and georeferenced global datasets. We analyse the long-term annual average pesticide residue and leaching rate of cumulative and individual substances. Model benchmark is provided

in the supplementary information (available online at stacks.iop.org/ERL/16/044051/mmedia) while the interpretation of our analyses is provided in section 3.

2. Methods

2.1. Pesticide application rates

We used PEST-CHEMGRIDS v1.0 [21] to obtain the global georeferenced annual pesticide application rates in year 2015 for nine cropping systems that include six dominant (i.e. alfalfa, corn, cotton, rice, soybean, and wheat) and three aggregated crops (i.e. vegetable and fruits, orchards and grapes, and other crops). Following the classification in the USGS Pesticide National Synthesis Project [22], the ‘vegetable and fruits’ cropping system includes legumes, roots and tubers, bush fruits, and herbaceous crops; the ‘orchards and grapes’ includes nuts, fruit trees, and vines; while the ‘other crops’ includes other cereals, oil crops, and fibre crops (refer to table 2 in Maggi *et al* [21] for a detailed classification).

PEST-CHEMGRIDS v1.0 provides the high and low annual application rates of 95 active substances that represent approximately 84% of the global pesticide mass used in 2015. In this study, we excluded calcium polysulfide, *Bacillus amyloliquefaciens*, and petroleum oil from our assessment due to insufficient data on their physicochemical properties. Hence, we assessed 92 substances using their median application rates.

2.2. Model description

We estimated the pesticide residue in soil using a mechanistic, spatially explicit and time-resolved model, solved within the general-purpose multi-phase and multi-component bioreactive transport simulator (BRTSim v4.0e [23]). The model explicitly describes the water, gas, and heat flow along a one-dimensional variably-saturated soil column, the transport of aqueous and gaseous chemical species, pesticide volatilisation, adsorption, and degradation.

BRTSim solves for the non-isothermal continuity and conservation laws using hybrid explicit-implicit numerical techniques within finite volumes (refer to the User Manual and Technical Guide for detailed descriptions [24]). The water flow is modelled using the Richards equation [25] in conjunction with the relative permeability-water potential-saturation relationships of the Brooks–Corey model [26]. The transport of aqueous species is modelled by the Darcy’s advection and the Fick’s diffusion equations. We excluded the advection of gaseous compounds, but their diffusion is explicitly described using the Fick’s law.

Pesticide volatilisation and adsorption are modelled using the mass action law [23], i.e.

$$K = \prod_R [X_R]^{-x_R} \cdot \prod_P [X_P]^{x_P}, \quad (1)$$

where $[X_R]$ and $[X_P]$ are the reactant and product concentrations, respectively, while x_R and x_P are their corresponding stoichiometric numbers. K is the Henry's constant for gas volatilisation. For adsorption, K is expressed as $K = K_{OC} \times SOC \times \frac{\rho_{soil}}{\theta}$, where K_{OC} is the soil organic carbon partition coefficient in ($\text{m}^3 \text{kg-C}^{-1}$), SOC is the soil organic carbon content in (kg-C kg-soil^{-1}), ρ_{soil} is the soil bulk density in (kg-soil m^{-3}), and θ is the soil water content in ($\text{m}^3 \text{m}^{-3}$).

We assumed that the degradation rate R of pesticides occurs by first-order kinetics and that this is moderated by biological activity, soil moisture content, temperature, pH, and organic carbon content, i.e.

$$R = f_B \times f_{pH} \times \frac{\ln 2}{DT_{50}} \times \frac{SOC}{SOC_{ref}} \times [X], \quad (2)$$

where $[X]$ and DT_{50} are the pesticide concentration and half-life, respectively. SOC_{ref} is the organic carbon content of the soil at which DT_{50} was estimated. Here, we used $SOC_{ref} = 0.02 \text{ kg-C kg-soil}^{-1}$, which is the global median. The control of pH on pesticide degradation is described by

$$f_{pH} = \frac{[H^+]}{[H^+] + K_{pH,L}} \times \frac{K_{pH,U}}{K_{pH,U} + [H^+]}, \quad (3)$$

with $[H^+]$ as the concentration of H^+ , and $K_{pH,L} = 1 \times 10^{-9} \text{ mol l}^{-1}$ and $K_{pH,U} = 1 \times 10^{-5} \text{ mol l}^{-1}$ representing the parameters of inhibition around the optimal value of pH 7. The function f_B describes the biological response to changes in soil saturation S and temperature T , expressed as $f_B = \min\{f(T), f(S) / \max\{f(S)\}\}$ [23] with

$$f(T) = \left(\frac{e^T}{e^{T_L} + e^T} \right)^{0.15} \times \left(\frac{e^{T_U}}{e^{T_U} + e^T} \right)^{0.35}, \quad (4)$$

$$f(S) = \left(\frac{S}{S_L + S} \right) \times \left(\frac{S_U}{S_U + S} \right), \quad (5)$$

where $T_L = 288.15 \text{ K}$ (i.e. $15 \text{ }^\circ\text{C}$) and $T_U = 313.15 \text{ K}$ (i.e. $40 \text{ }^\circ\text{C}$) are the lower and upper response temperatures that give an optimal rate in the range $T = 21 \text{ }^\circ\text{C}$ – $32 \text{ }^\circ\text{C}$ for mesophiles, while $S_L = S_U = 0.46$ are scalar parameters estimated from experimental data in Wickland and Neff (2008) [27] that give an optimal biological response at about $S = 0.5$.

The model was deployed on a three-dimensional grid resolved at $0.5^\circ \times 0.5^\circ$ resolution horizontally and extended vertically over two atmospheric and four soil layers. The atmospheric layers allow for water ponding and the exchange of heat and gas between soil and the atmosphere. The first three soil layers are variably saturated and extend down to 1 m depth in the root zone (RZ) with a thickness of 30 cm, 30 cm, and 40 cm, respectively. The last layer has a variable thickness that depends on the depth of either

the equilibrium water table or the bedrock and has a constant soil water saturation.

Pesticides were applied at the first soil layer with the annual applied mass distributed over the year following the crop calendar as in Maggi *et al* [28]. We considered a 40% loss of applied mass due to crop interception and wind drift after Trevisan *et al* [29]. Water and heat enter and leave the soil column through boundary fluxes that include rainfall, irrigation, crop evapotranspiration, and solar radiation (short and long waves). Incoming water fluxes and solar radiation were applied at the first soil layer, while the evapotranspiration was allocated over the soil profile according to the crop root distribution. The root distribution was calculated based on a negative exponential distribution function assuming that 99% of the root mass was above the maximum root depth.

We modelled 32 768 grid cells containing the nine cropping systems. The cells were selected based on the harvested area maps distributed along with PEST-CHEMGRIDS, which were originally produced by Monfreda *et al* [30]. Hence, we analysed in total 11.85 million km^2 of croplands.

2.3. Model input data

We fed the model with global georeferenced datasets described in table S1 of the supplementary information and we briefly list them below.

Soil physical properties, including soil texture (sand, silt, and clay fractions), bulk density, pH, and organic carbon content were obtained from the SoilGrids2.0 [31], while the soil porosity was obtained from SoilGrids1.0 [32]. The soil permeability, the pore volume distribution index and air-entry suction of the Brooks–Corey model, soil heat capacity, and heat conductivity were sourced from Dai *et al* [33]. The equilibrium water table was obtained from Fan *et al* [34], soil thickness from Pelletier *et al* [35], and soil residual liquid saturation from Zhang *et al* [36]. The model was initialised with the long-term average soil saturation and land surface temperature obtained from the CPC Soil Moisture dataset by NOAA/OAR/ESRL PSD, Boulder, Colorado, USA [37] and Menne *et al* [38], respectively.

We sourced the crop calendars from Sacks *et al* [39], which were used to calculate the daily pesticide application rates and irrigation volumes following a similar approach as in Maggi *et al* [28]. The crop water security indicator in Thenkabail *et al* [40] was used to determine if the agricultural land in a grid cell is irrigated.

We used the time series (from 1970 to 2017) of rainfall, atmospheric temperature, longwave and shortwave solar radiation, and potential evapotranspiration from the Climatic Research Unit time-series datasets [41] as the hydroclimatic forcing. The crop evapotranspiration was determined by multiplying the potential evapotranspiration with crop coefficient

k_c obtained from Allen *et al* [42]. The maximum root depth of each crop was also obtained from Allen *et al* [42].

The physicochemical properties of the 92 pesticides, including the molecular mass, Henry's law constant, half-life, adsorption affinity to organic carbon, and LC50 of earthworms were obtained from the Pesticide Properties DataBase [43].

2.4. Data analyses

We considered the top 30 cm of the soil (i.e. the first soil layer) as the TS, the top 1 m as the RZ, and the last soil layer as the BRZ. All simulations were run for 48 years and the outputs of the last 5 years were used for analyses described below.

The instantaneous residue M of pesticide X in the TS of cropping system j at time t was calculated as

$$M_X^j(t, \text{TS}) = \frac{C_X^j(t, \text{TS}) \times m_X \times V_w^j(t, \text{TS})}{M_s(\text{TS})}, \quad (6)$$

where C_X and m_X are the concentration (mol l^{-1}) and molecular mass of pesticide X , respectively, V_w is the water volume, and M_s is the soil mass. The total pesticide residue in cropping system j is the cumulative of all the 92 substances, i.e. $M_{\text{TOT}}^j(t, \text{TS}) = \sum_{X=1}^{92} M_X^j(t, \text{TS})$, while the total residue in a grid cell is the average residue of the nine cropping systems weighted by the crop harvested area A , i.e. $M_{\text{TOT}}(t, \text{TS}) = \sum \left(M_{\text{TOT}}^j(t, \text{TS}) \times A^j \right) / \sum_{j=1}^9 A^j$.

The mass of pesticides leached BRZ within a time window per unit area was calculated as

$$L_X^j = \frac{\Delta \left(C_X^j(t, \text{BRZ}) \times m_X \times V_w^j(t, \text{BRZ}) \right)}{\Delta t \times A^j}. \quad (7)$$

Similarly, the per-unit-area total leaching in a cropping system was determined as $L_{\text{TOT}}^j = \sum_{X=1}^{92} L_X^j$ and in a grid cell as $L_{\text{TOT}} = \sum \left(L_{\text{TOT}}^j \times A^j \right) / \sum_{j=1}^9 A^j$. Both residue and leachate included pesticides in aqueous and adsorbed phases.

In our analyses, major watersheds were identified based on AQUEDUCT-v2.1 [44]. The predicted no effect concentration of each pesticide is estimated as the LC50 of earthworms divided by a safety factor of 1000 as in the European Commission Risk Assessment guideline [45].

2.5. Model benchmark

We benchmarked our assessments against key environmental variables and the pesticide residues observed in European agricultural soils. Specifically, we benchmarked the modelled soil wetness, temperature, and pH in the TS (i.e. the top 30 cm) against values obtained from the CPC Soil Moisture dataset [37], the NOAA/NCEI land surface temperature

dataset [38], and the SoilGrids2.0 dataset [31]. We compared the modelled pesticide soil residues and the number of detectable substances in the TS against field observations reported in Silva *et al* [20]. Results of the model benchmarking are reported in text S1 of the supplementary information.

2.6. Sensitivity of variables and quality of estimation

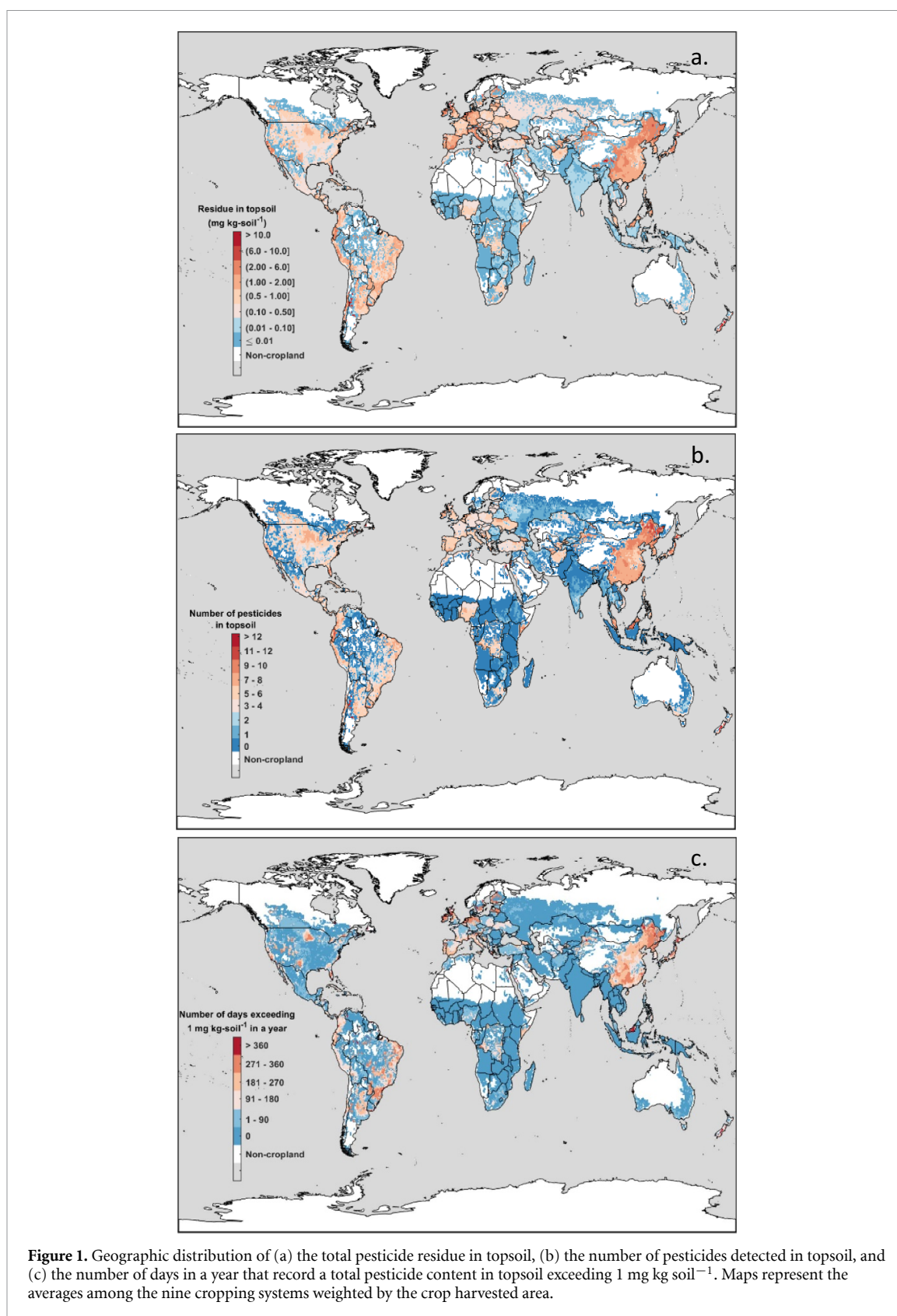
We used the AMAP index developed in la Cecilia *et al* [46] to assess the model sensitivity. AMAP quantifies the effect of a variable on the probability of a target output to exceed a threshold, with higher values signifying greater influences. The variables assessed include four boundary fluxes (i.e. pesticide application rate, rainfall, potential evapotranspiration, and net solar radiation) and seven soil properties (i.e. porosity, bulk density, permeability, pore volume distribution index, air-entry suction, pH, and organic carbon content). The target model outputs are the pesticide residue in the TS and leaching BRZ with corresponding thresholds of $0.01 \text{ mg kg-soil}^{-1}$ and $1 \text{ mg m}^{-2} \text{ yr}^{-1}$, respectively.

To measure the quality of our estimates (QI), we integrated the data quality QI_{APR} of PEST-CHEMGRIDS v1.0 and the quality of benchmarking, i.e. $QI = (3 \cdot QI_{\text{APR}} + QI_{\text{S}} + QI_{\text{T}} + QI_{\text{pH}}) / 6$. The overall quality of the application rate data QI_{APR} was calculated as the average over all crops and all substances. The qualities of soil saturation (QI_{S}), temperature (QI_{T}), and pH (QI_{pH}) were calculated as the normalised absolute errors in soil saturation, temperature, and pH against global datasets described above. All quality indices are bounded between 0 and 1 such that $QI = 0$ and $QI = 1$ signify low and high data quality, respectively.

3. Results and discussion

3.1. Pesticide residue in topsoil

Globally, the TS in croplands has an average total pesticide content of $0.62 \text{ mg kg-soil}^{-1}$ and a 95th percentile of $2.12 \text{ mg kg-soil}^{-1}$, with residue heterogeneously distributed across geographic regions (figure 1(a)[47]). Specifically, the five countries with the highest average pesticide residue are New Zealand ($30.7 \text{ mg kg-soil}^{-1}$), United Kingdom ($14.4 \text{ mg kg-soil}^{-1}$), Cyprus ($5.2 \text{ mg kg-soil}^{-1}$), The Netherlands ($3.6 \text{ mg kg-soil}^{-1}$), and Japan ($2.7 \text{ mg kg-soil}^{-1}$, table S2, supplementary information). The majority of croplands in Northern, Southern, and Western Europe have pesticide residues higher than the global average (i.e. about 76.8%), especially in Italy, Spain, Germany, and United Kingdom, while residues in Eastern Europe are relatively lower (figure 1(a)). We estimated that pesticide residue in about 1.88 million km^2 of croplands exceeded $1 \text{ mg kg-soil}^{-1}$ for more than 180 d in a year (figure 1(c)). These regions are mainly located in China (0.83 million km^2), Brazil



(0.23 million km^2), and the United States (0.17 million km^2 , table S2, supplementary information). Our analysis shows that more than 90% of pesticide residue remains in the adsorbed phase, especially in regions with high residue contents including China,

Europe, and North America (figure S6, supplementary information), suggesting that sorption kinetics are important mechanisms that determined the pesticide accumulation in soil. Croplands in Africa, South and Southeast Asia, and Australia have relatively low

pesticide residue, with an exception in a few countries such as Malaysia and the Philippines (figure 1(a)).

We counted the number of pesticides with a residue greater than the typical laboratory measuring limit (i.e. $0.01 \text{ mg kg-soil}^{-1}$) and found that 70% of global croplands (i.e. about 8.31 million km^2) have multiple pesticide residues in the TS (figure 1(b)[47]). More than ten pesticide residues are detected in 16 major watersheds (figure S7, supplementary information), including Amur and Huang He (China), Colorado and Negro (Argentina), Rajang (Malaysia), Amazonas and Esmeraldas (Ecuador), Nile (Egypt), Ishikari (Japan), and Clutha (New Zealand). Croplands in Southern and Western Europe have, on average, four to five detectable pesticide residues despite the high total pesticide contents found.

Among the nine cropping systems, ‘orchards and grapes’ has the highest average total residue (i.e. $2.35 \text{ mg kg-soil}^{-1}$) with the highest number of detectable pesticides (figures S8(a) and (b), supplementary information). Specifically, ‘orchards and grapes’ in Japan and Chile have more than 14 pesticide residues that summed up to more than $26 \text{ mg kg-soil}^{-1}$. ‘Vegetable and fruits’ also records high content of pesticide mixtures, especially in Malaysia and New Zealand, which have more than $24 \text{ mg kg-soil}^{-1}$ of residue. Croplands for corn and soybean have comparable pesticide residue (i.e. a global average of about $1 \text{ mg kg-soil}^{-1}$), while wheat fields have the lowest (i.e. a global average of $0.12 \text{ mg kg-soil}^{-1}$).

The five most frequently detected pesticides in the TS are glyphosate, pendimethalin, chlorpyrifos, paraquat, and chlorothalonil (figure S7, supplementary information). Although their global average residue is relatively low (i.e. $<0.4 \text{ mg kg-soil}^{-1}$), some major watersheds are exposed to residue exceeding the predicted no-effect concentrations for earthworms (figure 2(a)). In particular, watersheds in Argentina and Chile are exposed to chlorpyrifos residue of one order of magnitude higher than the no-effect threshold, while several watersheds in Asia and South America have average chlorothalonil residue exceeding the no-effect threshold by more than three times, such as in Lake Titicaca, Brahmaputra, Majes, Salween, and Rajang (figure 2(a)). Among all pesticides assessed, the global 95th percentiles of 13 pesticide residues exceeded the no-effect threshold (figure S9(a), supplementary information). Bromoxynil, phorate, and thiophanate-methyl are among the most concerning pesticides as their global average residues are more than two factors higher than the no-effect threshold (figure S9(a), supplementary information).

3.2. Pesticides leached below the root zone

Every year, approximately 0.2 million tonnes of pesticides leach BRZ globally, corresponding to about 5.6% of the applied mass. The geographic

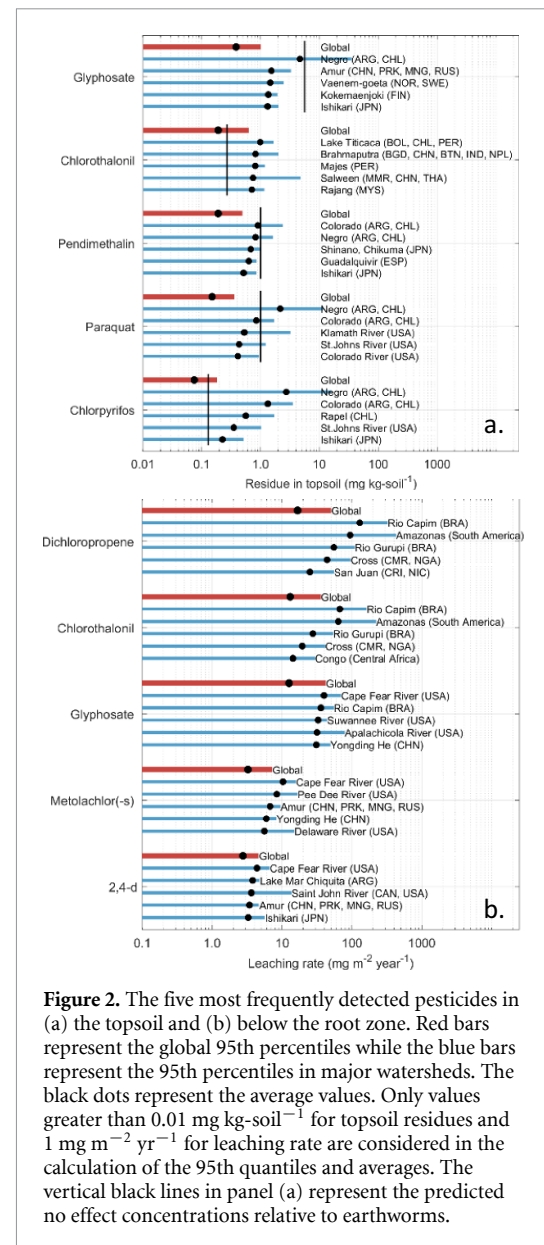


Figure 2. The five most frequently detected pesticides in (a) the topsoil and (b) below the root zone. Red bars represent the global 95th percentiles while the blue bars represent the 95th percentiles in major watersheds. The black dots represent the average values. Only values greater than $0.01 \text{ mg kg-soil}^{-1}$ for topsoil residues and $1 \text{ mg m}^{-2} \text{ yr}^{-1}$ for leaching rate are considered in the calculation of the 95th quantiles and averages. The vertical black lines in panel (a) represent the predicted no effect concentrations relative to earthworms.

distribution of high leaching rate areas are substantially different from regions with high pesticide residue (figure 3(a) [47]). The leaching rate in equatorial regions is especially high, such as in Amazonas, Niger, and Congo. In particular, approximately $10\,600 \text{ km}^2$ in the Amazonas watershed leach more than $100 \text{ mg m}^{-2} \text{ yr}^{-1}$ and, in total, 2800 tonnes leach BRZ of the watershed every year. In Asia, leaching is particularly high in parts of China, Japan, and the Philippines. Leaching greater than $200 \text{ mg m}^{-2} \text{ yr}^{-1}$ are observed in the southern regions of China, including Xi Jiang, Min Jiang, and Dong Jiang. A total of about 80 000 tonnes leach BRZs in China every year, contributing to 40% of the global pesticide leachate. Although European croplands see high pesticide residue in the TS, their leaching is generally low, with high rates mainly observed in the alpine regions between North Italy and Austria (figure 3(a)).

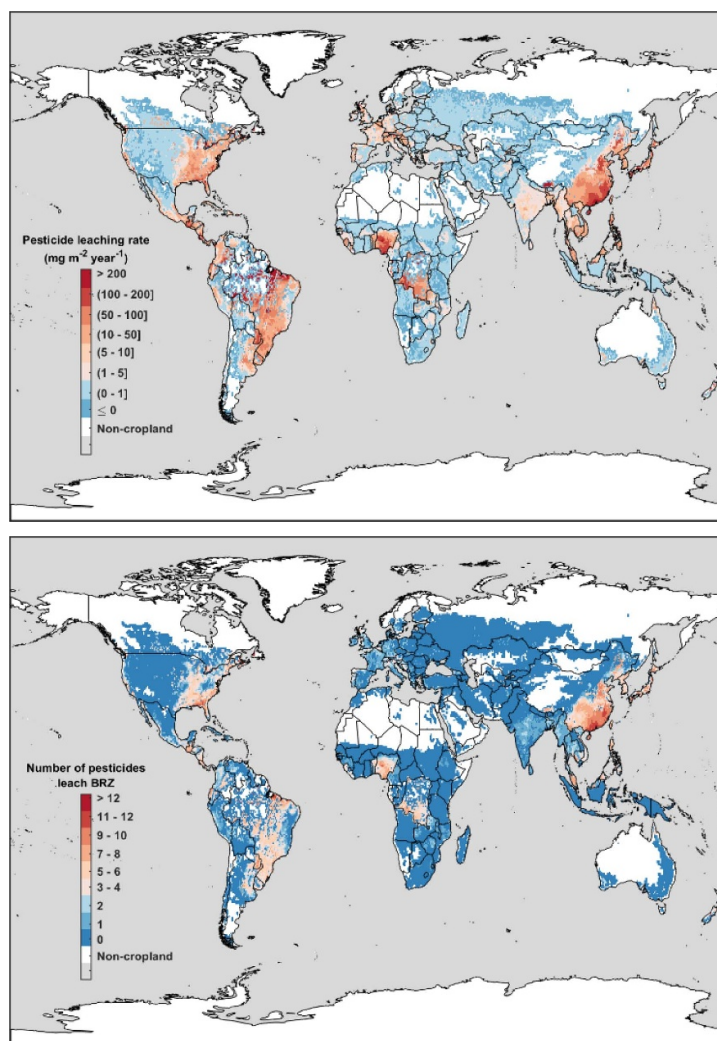


Figure 3. Geographic distribution of (a) the total pesticide leaching rate, and (b) the number of pesticides leaching below the root zone (BRZ) at a rate greater than $1 \text{ mg m}^{-2} \text{ yr}^{-1}$. Maps represent the averages among the nine cropping systems weighted by the crop harvested area.

In figure 3(b) [47], we counted the number of pesticides that exceeded a leaching rate of $1 \text{ mg m}^{-2} \text{ yr}^{-1}$, which corresponds to the regulatory limit of groundwater contamination stated by the European Commission (i.e. $0.1 \mu\text{g l}^{-1}$ [48]) after assuming that a shallow aquifer is located BRZ with a thickness of 23 m and a soil porosity of 0.43 (which are the global median values derived from SoilGrids1.0 [32], Fan *et al* [34], and Pelletier *et al* [35]). Leaching of more than two pesticides is mainly observed in the south-eastern of the United States, Brazil, Nigeria, Congo, China, Japan, Malaysia, and the Philippines. In particular, approximately 9% of croplands in China (i.e. about 0.13 million km^2 , mainly in Yangtze River, Xi Jiang, and Min Jiang) are subject to leaching of more than ten pesticides (figure 3(b)).

Globally, ‘vegetable and fruits’ cropping system has the highest average leaching rate (i.e. $53 \text{ mg m}^{-2} \text{ yr}^{-1}$) and the greatest number of pesticides that exceeded $1 \text{ mg m}^{-2} \text{ yr}^{-1}$, especially in

Guatemala where more than ten pesticides exceeded the threshold (figures S8(c) and (d), supplementary information). On average, more than 10% of the applied mass in soybean and rice fields leaches BRZ. Wheat croplands have the lowest leaching rate.

Dichloropropene, chlorothalonil, glyphosate, metolachlor, and 2,4-d are the most frequently detected pesticides to exceed the leaching threshold (figure 2(b)), among which glyphosate contributes to the greatest total mass leached BRZ (i.e. about 50 000 tonnes yr^{-1}). In addition to the pesticides mentioned above, bromoxynil, metam, metam potassium, propanil, and quinclorac also have high leaching rates, with the global 95th percentile values exceeding $20 \text{ mg m}^{-2} \text{ yr}^{-1}$ (figure S9(b), supplementary information).

3.3. Sensitivity of variables and quality of estimates

Pesticide application rate is the most influential variable for both the TS residue and leaching BRZ to exceed the defined thresholds of $0.01 \text{ mg kg-soil}^{-1}$

and $1 \text{ mg m}^{-2} \text{ yr}^{-1}$, respectively (figure S10, supplementary information). In general, model outputs are also sensitivity to variations in boundary fluxes. Relative to TS residues, potential evapotranspiration and net solar radiation have a higher control over the probability of exceeding the threshold than soil properties (figure S10(a), supplementary information), whereas the leaching rate is impacted by the rainfall amount and soil pH (figure S10(b), supplementary information).

Since pesticide application rate has a major impact on the model outputs, we propagated the uncertainties in PEST-CHEMGRIDS toward quantifying the data quality of our model estimates as described in section 2.6. A high QI index signifies good quality of estimates. The distribution of the QI index is right-skewed with a median at 0.7 (figure S11(b), supplementary information). Less than 8% of the grid cells have an QI index smaller than 0.5 and are mainly located in Brazil, Saudi Arabia, Iran, and Western regions of China (figure S11(a), supplementary information).

4. Discussion

Our assessment extends the study in Silva *et al* [20] and addresses the soil contamination by pesticide mixtures at the global scale. We showed that the simultaneous existence of multiple pesticides is very much a normality in agricultural soil worldwide and, in some regions, the cumulative residues can exceed $10 \text{ mg kg-soil}^{-1}$.

More strikingly, residues of many pesticides (individually) exceeded the no-effect concentrations for earthworms, but we do not exactly know how synergistic effects of these mixtures affect earthworms. Among pesticides most frequently detected in the TS, chlorpyrifos, chlorothalonil, and dichloropropene have particularly high toxicities to soil biota and their residue exceeded the no-effect threshold in some areas. Previous studies show that the inhibitory effects of chlorpyrifos on soil microbial activity and population were significantly amplified in the presence of chlorothalonil [49, 50], while noting that chlorpyrifos and chlorothalonil residues frequently coexist, especially in orchards and grapes cropping system. Yet, only less than 102 jurisdictions (not countries) worldwide have promulgated regulatory guidance values relative to soil residues for these compounds (i.e. 102 for chlorpyrifos, 75 for dichloropropene, and 67 for chlorothalonil [15]) with some guideline values being a few times higher than the LC50 of earthworms. For example, the guideline value of chlorpyrifos in Michigan, USA, is $11\,000 \text{ mg kg-soil}^{-1}$ [15], which is nearly two orders of magnitude greater than the LC50 of earthworms (i.e. $129 \text{ mg kg-soil}^{-1}$ [43]). In addition to earthworms, pesticides can have adverse

effects on soil microbial communities such as nitrifying bacteria and archaea that are responsible for transforming nitrogen into forms available for microbial and plant uptake [10]. Some pesticides can also increase the rate of ammonification [51], i.e. the fast conversion of amino acids into ammonium ions that are more prone to leaching, leading to nitrogen loss. More dreadfully, pesticides can reduce the symbiotic efficiency of nitrogen-fixing rhizobia bacteria [52], decreasing the ability for legume crops to replenish nitrogen in the soil. Hence, pesticide contamination may result in soil fertility loss. However, a comprehensive assessment of pesticide contamination risk on soil biota is currently unachievable owing to the lack of ecotoxicological data specific to each pesticide substance and soil biota species.

We also estimated that nearly 6% of the applied pesticides leach BRZ, where their degradation rates are substantially low. Pesticides may persist and accumulate in the deep soil and potentially contaminate the groundwater. Over the years, numerous pesticides were detected in the groundwater across continents, including the United States [53], Canada [54], Europe [55], and China [56], posing threats to the safety of drinking water supplies.

Our assessment suggests the need to establish a coherent regulatory guideline for pesticide contamination in soil that is in concordance with environmental and ecotoxicological studies to protect soil functions while acknowledging that there are many knowledge gaps yet to be filled. The combinatorial effects of multiple pesticides on soil biota, nutrient cycles, and crop uptake and metabolism are still not well understood, leading to high uncertainties on how pesticide residue affects soil fertility and crop yield. In a warming world, climate change adds on another layer of complexity in illustrating the role of pesticides in crop production, whether they are the protection or threats to the food security of the future.

Our model assessment provides a starting point to address the knowledge gaps. It identifies the pesticides, cropping systems, and geographic regions that require research investment and tailored strategies. Yet, we acknowledge the numerical and model limitations embedded in our estimates. We did not account for the transport and fate of the metabolites, which can retain the toxicity of parent substances and persist in the environment, and therefore we may underestimate the extent of contamination. Our modeling did not explicitly include surface runoff and soil erosion, the processes that can enhance contaminant dispersion. We conducted a parameter sensitivity screening by treating each grid cell on the map as an individual replica. This may not be the most ideal method for uncertainty quantification, but a comprehensive global sensitivity analysis is not feasible due to high computational cost, i.e. one full model run for

the work presented here consumed more than 30 000 CPU hours.

5. Conclusions

We presented an estimate of global soil contamination by pesticide mixtures. Globally, the pesticide residue in the TS of about 1.88 million km² of croplands exceeded a concentration of 1 mg kg-soil⁻¹ for more than 180 d in a year, while approximately 5.6% of the applied pesticides leach BRZ. High residue in the TS and leaching BRZ are estimated for ‘orchards and grapes’ and ‘vegetable and fruits’ cropping systems, whereas wheat fields have the lowest pesticide contamination. Croplands in parts of Europe, South America, China, and Japan have high pesticide residue with more than 10 pesticides detected in 16 major watersheds. Leaching is particularly high in equatorial and southern regions of China. Among the 92 pesticides assessed, glyphosate and chlorothalonil are the most frequently detected in the TS and to leach BRZ. Our assessment urges for more research efforts in understanding pesticide impacts on soil health and functions.

Data availability statement

The data that support the findings of this study are openly available at the following URL/DOI: <https://doi.org/10.6084/m9.figshare.12966323>. Data will be available from 01 February 2021.

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Code availability

The BRTSim software can be freely downloaded from <https://sites.google.com/site/thebrtsimproject>. An example of input files required to run the model for this work can be downloaded via *figshare* at doi: <https://doi.org/10.6084/m9.figshare.12966323>.

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References

- [1] FAOSTAT 2020 Database collection of the food and agriculture organization of the United Nations FAOSTAT Food and Agriculture Organization of the United Nations (available at: www.fao.org/faostat/en/#data) (Accessed 11 August 2020)
- [2] European Commission 2016 PLANT EU pesticides database (available at: <http://ec.europa.eu/food/plant/pesticides/eu-pesticides-database/>) (Accessed 2019)
- [3] Landrigan P J *et al* 2017 The Lancet Commission on pollution and health *The Lancet* **391** 462–512
- [4] Sannino F and Gianfreda L 2001 Pesticide influence on soil enzymatic activities *Chemosphere* **45** 417–25
- [5] Puglisi E 2012 Response of microbial organisms (aquatic and terrestrial) to pesticides *EFSA Support. Publ.* **9** 359E
- [6] Karas P A *et al* 2018 Assessment of the impact of three pesticides on microbial dynamics and functions in a lab-to-field experimental approach *Sci. Total Environ.* **637** 636–46
- [7] Thiour-Mauprivez C, Martin-Laurent F, Calvayrac C and Barthelmebs L 2019 Effects of herbicide on non-target microorganisms: towards a new class of biomarkers? *Sci. Total Environ.* **684** 314–25
- [8] Singh B K, Walker A and Wright D J 2002 Persistence of chlorpyrifos, fenamiphos, chlorothalonil, and pendimethalin in soil and their effects on soil microbial characteristics *Bull. Environ. Contam. Toxicol.* **69** 181–8
- [9] Wan R, Wang Z and Xie S 2014 Dynamics of communities of bacteria and ammonia-oxidizing microorganisms in response to simazine attenuation in agricultural soil *Sci. Total Environ.* **472** 502–8
- [10] Feld L, Hjelmsø M H, Nielsen M S, Jacobsen A D, Rønn R, Ekelund F, Krogh P H, Strobel B W and Jacobsen C S 2015 Pesticide side effects in an agricultural soil ecosystem as measured by amoA expression quantification and bacterial diversity changes *PLoS One* **10** e0126080
- [11] Pelosi C, Barot S, Capowiez Y, Hedde M and Vandenbulcke F 2014 Pesticides and earthworms. A review *Agron. Sustain. Dev.* **34** 199–228
- [12] Imfeld G and Vuilleumier S 2012 Measuring the effects of pesticides on bacterial communities in soil: a critical review *Eur. J. Soil Biol.* **49** 22–30
- [13] Yang G, Chen C, Wang Y, Peng Q, Zhao H, Guo D, Wang Q and Qian Y 2017 Mixture toxicity of four commonly used pesticides at different effect levels to the epigeic earthworm, *Eisenia fetida* *Ecotoxicol. Environ. Saf.* **142** 29–39
- [14] van Hoesel W *et al* 2017 Single and combined effects of pesticide seed dressings and herbicides on earthworms, soil microorganisms, and litter decomposition *Front. Plant Sci.* **8** 215
- [15] Jennings A A and Li Z 2014 Scope of the worldwide effort to regulate pesticide contamination in surface soils *J. Environ. Manage.* **146** 420–43
- [16] Römbke J, Bernard J and Martin-Laurent F 2018 Standard methods for the assessment of structural and functional diversity of soil organisms: a review *Integr. Environ. Assess. Manage.* **14** 463–79
- [17] Tang W, Wang D, Wang J, Wu Z, Li L, Huang M, Xu S and Yan D 2018 Pyrethroid pesticide residues in the global environment: an overview *Chemosphere* **191** 990–1007
- [18] Hvězdová M *et al* 2018 Currently and recently used pesticides in Central European arable soils *Sci. Total Environ.* **613** 361–70
- [19] Fernandes C L F, Volcão L M, Ramires P F, de Moura R R and Júnior F M R D S 2020 Distribution of pesticides in

- agricultural and urban soils of Brazil: a critical review *Environ. Sci.: Process. Impacts* **22** 256–70
- [20] Silva V, Mol H G, Zomer P, Tienstra M, Ritsema C J and Geissen V 2019 Pesticide residues in European agricultural soils—a hidden reality unfolded *Sci. Total Environ.* **653** 1532–45
- [21] Maggi F, Tang F H, la Cecilia D and McBratney A 2019 PEST-CHEMGRIDS, global gridded maps of the top 20 crop-specific pesticide application rates from 2015 to 2025 *Sci. Data* **6** 1–20
- [22] Baker N T 2017 Estimated annual agricultural pesticide use by major crop or crop group for states of the conterminous United States, 1992–2016 Estimated Annual Agricultural Pesticide Use U.S. Department of the Interior and U.S. Geological Survey (available at: <https://water.usgs.gov/nawqa/pnsp/usage/maps/index.php>) (Accessed 2020)
- [23] Maggi F 2019 BRTSim, a general-purpose computational solver for hydrological, biogeochemical, and ecosystem dynamics (<https://doi.org/arXiv:1903.07015>)
- [24] Maggi F 2019 BRTSim v4.0 release e, A general-purpose multiphase and multispecies computational solver for biogeochemical reaction-advection-dispersion processes in porous and non-porous media *User Manual and Technical Guide* (Sydney: The University of Sydney) p 85 (<https://sites.google.com/site/thebrtsimproject/download>)
- [25] Richards L A 1931 Capillary conduction of liquids through porous mediums *Physics* **1** 318–33
- [26] Brooks R H and Corey A T 1966 Properties of porous media affecting fluid flow *Journal of the irrigation and drainage division* **92** 61–88
- [27] Wickland K P and Neff J C 2008 Decomposition of soil organic matter from boreal black spruce forest: environmental and chemical controls *Biogeochemistry* **87** 29–47
- [28] Maggi F, la Cecilia D, Tang F H and McBratney A 2020 The global environmental hazard of glyphosate use *Sci. Total Environ.* **717** 137167
- [29] Trevisan M, di Guardo A and Balderacchi M 2009 An environmental indicator to drive sustainable pest management practices *Environ. Model. Softw.* **24** 994–1002
- [30] Monfreda C, Ramankutty N and Foley J A 2008 Farming the planet: 2. Geographic distribution of crop areas, yields, physiological types, and net primary production in the year 2000 *Glob. Biogeochem. Cycles* **22** GB1022
- [31] de Sousa L M, Poggio L, Batjes N H, Heuvelink G B M, Kempen B, Riberio E and Rossiter D 2020 SoilGrids 2.0: producing quality-assessed soil information for the globe (<https://doi.org/10.5194/soil-2020-65>)
- [32] Hengl T *et al* 2017 SoilGrids250m: global gridded soil information based on machine learning *PLoS One* **12** e0169748
- [33] Dai Y, Xin Q, Wei N, Zhang Y, Shangguan W, Yuan H, Zhang S, Liu S and Lu X 2019 A global high-resolution data set of soil hydraulic and thermal properties for land surface modeling *J. Adv. Model. Earth Syst.* **11** 2996–3023
- [34] Fan Y, Li H and Miguez-Macho G 2013 Global patterns of groundwater table depth *Science* **339** 940–3
- [35] Pelletier J D, Broxton P D, Hazenberg P, Zeng X, Troch P A, Niu G, Williams Z C, Brunke M A and Gochis D 2016 Global 1-km gridded thickness of soil, regolith, and sedimentary deposit layers ORNL DAAC (Oak Ridge, TN: Oak Ridge National Laboratory) (<https://doi.org/10.3334/ORNLDAAC/1304>)
- [36] Zhang Y, Schaap M G and Zha Y 2018 A high-resolution global map of soil hydraulic properties produced by a hierarchical parameterization of a physically based water retention model *Water Resour. Res.* **54** 9774–90
- [37] Fan Y and van den Dool H 2004 Climate prediction center global monthly soil moisture data set at 0.5 resolution for 1948 to present *J. Geophys. Res.: Atmos.* **109** D10102
- [38] Menne M J, Durre I, Vose R S, Gleason B E and Houston T G 2012 An overview of the global historical climatology network-daily database *J. Atmos. Ocean. Technol.* **29** 897–910
- [39] Sacks W J, Deryng D, Foley J A and Ramankutty N 2010 Crop planting dates: an analysis of global patterns *Glob. Ecol. Biogeogr.* **19** 607–20
- [40] Thenkabail P S *et al* 2016 NASA making earth system data records for use in research environments (MEaSURES) global food security support analysis data (GFSAD) crop dominance 2010 global 1 km V001. Monograph NASA EOSDIS Land Processes DAAC (South Dakota, USA) (<http://oar.icrisat.org/10982/>)
- [41] Harris I, Osborn T J, Jones P and Lister D 2020 Version 4 of the CRU TS monthly high-resolution gridded multivariate climate dataset *Sci. Data* **7** 1–18
- [42] Allen R G, Pereira L S, Raes D and Smith M 1998 Crop evapotranspiration: guidelines for computing crop water requirements—FAO irrigation and drainage paper 56 (Rome: Food and Agriculture Organisation of the United Nations (FAO))
- [43] Lewis K A, Tzilivakis J, Warner D J and Green A 2016 An international database for pesticide risk assessments and management *Hum. Ecol. Risk Assess.* **22** 1050–64
- [44] Gassert F, Luck M, Landis M, Reig P and Shiao T 2014 Aqueduct global maps 2.1: constructing decision-relevant global water risk indicators Washington, DC: World Resources Institute (<https://www.wri.org/publication/aqueduct-global-maps-21-indicators>)
- [45] European Commission 2003 Technical Guidance Document on Risk Assessment in support of Commission Directive 93/67/EEC on Risk Assessment for new notified substances (Italy: Institute for Health and Consumer Protection)
- [46] la Cecilia D, Porta G M, Tang F H, Riva M and Maggi F 2020 Probabilistic indicators for soil and groundwater contamination risk assessment *Ecol. Indic.* **115** 106424
- [47] Tang F H M and Maggi F 2021 Pesticide mixtures in soil: a global outlook figshare (<https://doi.org/10.6084/m9.figshare.12966323>)
- [48] European Commission 2006 Directive 2006/118/EC of the European Parliament and of the council of 12 December 2006 on the protection of groundwater against pollution and deterioration *Official Journal of the European Union* **L 372** 19–31 (<http://data.europa.eu/eli/dir/2006/118/oj>)
- [49] Singh B K, Walker A and Wright D J 2002 Degradation of chlorpyrifos, fenamiphos, and chlorothalonil alone and in combination and their effects on soil microbial activity *Environ. Toxicol. Chem.* **21** 2600–5
- [50] Chu X, Fang H, Pan X, Wang X, Shan M, Feng B and Yu Y 2008 Degradation of chlorpyrifos alone and in combination with chlorothalonil and their effects on soil microbial populations *J. Environ. Sci.* **20** 464–9
- [51] Wainwright M 1978 A review of the effects of pesticides on microbial activity in soils *J. Soil Sci.* **29** 287–98
- [52] Fox J E, Gullledge J, Engelhaupt E, Burow M E and McLachlan J A 2007 Pesticides reduce symbiotic efficiency of nitrogen-fixing rhizobia and host plants *Proc. Natl Acad. Sci.* **104** 10282–7
- [53] Toccalino P L, Gilliom R J, Lindsey B D and Rupert M G 2014 Pesticides in groundwater of the United States: decadal-scale changes, 1993–2011 *Groundwater* **52** 112–25
- [54] Munira S, Farenhorst A, Sapkota K, Nilsson D and Sheedy C 2018 Auxin herbicides and pesticide mixtures in groundwater of a Canadian prairie province *J. Environ. Qual.* **47** 1462–7
- [55] Leistra M and Boesten J J T I 1989 Pesticide contamination of groundwater in western Europe *Agric. Ecosyst. Environ.* **26** 369–89
- [56] Kong L, Kadokami K, Duong H T and Chau H T C 2016 Screening of 1300 organic micro-pollutants in groundwater from Beijing and Tianjin, North China *Chemosphere* **165** 221–30

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