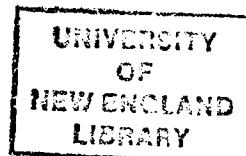


Identification and Characterisation of Dominant Ivermectin
Resistant Genes in *Caenorhabditis elegans*.

Peter William Hunt B. Rur. Sci. (Hons)
(University of New England).



Submitted: September, 1995

*A thesis submitted for the degree of Doctor of Philosophy of
the University of New England.*

ACKNOWLEDGMENTS.

During the three and a half years in which this thesis, and the experimental work described within it, have been undertaken, many people have provided assistance, guidance, comments and moral support.

Firstly, I would like to thank Dr. Warwick Grant as my principal supervisor for expert guidance, comments and assistance. Special thanks must be awarded to Warwick for having the foresight to re-introduce *C. elegans* research to Australia, thus providing me with an opportunity to work with this noble creature. My other supervisors, Dr. Margaret Katz and Dr. Brian Cheetham have also provided me with helpful comments and advice over the period of my candidature and have provided invaluable help during thesis preparation.

Invaluable assistance was also provided by Dr. Leo LeJambre and Dr. Carl Johnson, who made important contributions to my understanding of parasitic nematodes and ivermectin resistance respectively. I would also like to thank all workers in the molecular parasitology laboratory, Warwick Grant, Leo LeJambre, Ian Lenane, Lisa Mascord, Gill Whitington and Michelle Wooster for helpful discussions, advice, technical help and comradeship.

Thanks also to the *C. elegans* research community for the support network that is provided for all workers in the field. Especially I would like to thank Drs L. Avery, D. Baillie, A. Coulson, R. Herman, J. Hodgkin, C. Johnson, D. Riddle, T. Stiernagle and J. Thomas for helpful communications with myself (or with Warwick on my behalf) and/or for supplying strains.

I also acknowledge the support of CSIRO Division of Animal Health, the Australian Wool Corporation and the University of New England, who provided facilities and financial support.

Finally I would like to thank my friends and family for being understanding and supportive throughout my PhD, and especially my wife Raelene who has endured the last three and a half years, providing sympathy, encouragement and love.

TABLE OF CONTENTS

LIST OF FIGURES.....	x
LIST OF TABLES.....	xi
ABSTRACT.....	xiii
AIMS.....	1
CHAPTER ONE.....	2
1.1. The mode of action of ivermectin.....	4
1.1.1. Behavioural observations.....	7
1.1.2. Biochemistry.....	9
1.1.3. Electrophysiology.....	10
1.1.4. Molecular Biology of the IVM receptor from <i>Caenorhabditis elegans</i>	12
1.1.5. Mode of action of IVM in nematodes.....	13
1.2. Genetics of ivermectin resistance in <i>Caenorhabditis</i> <i>elegans</i> and intestinal parasites of sheep.....	15
1.3. Sensory structures and their functions in <i>Caenorhabditis</i> <i>elegans</i>	19
1.3.1. Anterior sensilla, phasmids and deirids.....	20
1.3.1.1. Amphids.....	22
1.3.1.2. Inner Labial Sensilla	33
1.3.1.3. Outer Labial and Cephalic sensilla.....	34
1.3.1.4. Phasmids and deirids.....	35
1.3.2. Touch receptors.....	36
1.3.3. Sensory structures of the male tail.....	39
1.3.4. Pharyngeal sensory neurons.....	41
1.3.5. Other putative sensory neurons.....	41
1.3.6. Conclusions	43
CHAPTER TWO.....	44
2.1. INTRODUCTION.....	45
2.2. METHODS.....	47
2.2.1. Determination of resistance status	47
2.2.2. Preparation of agar plates containing IVM.....	47
2.2.3. Amphid neuron staining of <i>Haemonchus</i> <i>contortus</i> strains.....	48
2.2.3.1. Staining of L1 and L2 larvae with DiO.....	48
2.2.3.2. Staining of L3 <i>H. contortus</i> with FITC.....	49
2.2.3.3. Staining of Adult <i>Haemonchus</i> with DiO.....	49
2.2.4. Electron Microscopy of amphids of L2 <i>Haemonchus contortus</i> larvae.....	49
2.2.5. Analysis of IVM resistant <i>C. elegans</i> mutants for additional phenotypes.....	50
2.2.5.1. Staining of amphid neurons of IVM resistant strains	50
2.2.5.2. Analysis of <i>che-3(nr5)</i> for dauer formation.....	50
2.2.5.3. Complementation testing of <i>che-3(nr5)</i> with <i>che-3(e1124)</i> and <i>che-13(e1805)</i> for IVM resistance.....	50
2.2.5.4. Growth of Df mutants at 25°C.....	51

2.2.6. Chemosensory behaviour of IVM sensitive <i>C. elegans</i> strains on IVM.....	51
2.3. RESULTS	
2.3.1. Resistance status of mutant strains.....	51
2.3.1.1. Strains carrying mutations in <i>unc</i> genes and other neuronal defective mutants.....	52
2.3.1.2. Neurotransmitter defective strains.....	52
2.3.1.3. Dyf mutations confer IVM resistance.....	55
2.3.1.4. Mutants of <i>C. elegans</i> which are Supersensitive to IVM.....	57
2.3.1.5. Some mutants with defects in head morphology are slightly resistant to IVM.....	59
2.3.2. Staining of <i>Haemonchus contortus</i> amphid neurons.....	61
2.3.3. Electron microscopy of IVM resistant and sensitive <i>Haemonchus contortus</i>	62
2.3.4. A number of additional phenotypes are associated with IVM resistance.....	66
2.3.4.1. Mutants selected for IVM resistance are also Dyf.....	66
2.3.4.2. <i>che-3(nr5)</i> is dauer formation defective.....	66
2.3.4.3. IVM resistance of <i>dyf</i> mutants is temperature sensitive.....	66
2.3.5. <i>C. elegans</i> sensitive to IVM show disorientation when placed on IVM media.....	68
2.4. DISCUSSION	
2.4.1. Resistance status of mutant strains.....	70
2.4.1.1. IVM resistance of mutants with neuronal defects is associated with defects in amphid neuron sensory cilia.....	70
2.4.1.2. IVM sensitivity is influenced by exposure of amphid neuron sensory cilia to the external environment.....	71
2.4.1.3. Mutants selected for IVM resistance are alleles of Dyf loci.....	74
2.4.1.4. Supersensitivity to IVM is conferred by a range of mutations.....	76
2.4.2. Staining of <i>Haemonchus contortus</i> amphid neurons and electron microscopy.....	78
2.4.2.1. IVM resistance and amphid defects in <i>H. contortus</i>	78
2.4.2.2. Comparisons of amphid morphology between developmental stages and between species.....	79
2.4.3. Ivermectin acts to disorient sensitive nematodes.....	82
CHAPTER THREE	85
3.1. INTRODUCTION	86
3.2. METHODS	91
3.2.1. Mutagenesis screening for mutants carrying dominant IVM resistant mutations.....	91
3.2.2. Confirmation of dominance.....	92

3.2.3. Construction of new strains from the strains isolated in the mutagenesis experiments.....	93
3.2.3.1. Outcrossing dominant IVM resistant strains.....	93
3.2.3.2. Construction of marked IVM resistant strains.....	93
3.2.4. IVM dose response of homozygotes and heterozygotes.....	94
3.2.4.1. Determination of resistance of homozygotes to different IVM concentrations.....	94
3.2.4.2. Determination of resistance of heterozygotes to different IVM concentrations.....	94
3.2.4.3. Generation of heterozygotes for dose response experiments.....	95
3.2.5. Determination of Dyf phenotype of homozygotes and heterozygotes.....	96
3.2.5.1. Staining of amphid neurons of IVM resistant strains.....	96
3.2.5.2. DiO Plates	96
3.2.5.3. Generation of heterozygotes for staining.....	96
3.2.6. Linkage analysis of dominant strains.....	97
3.2.6.1. Sex linkage.....	97
3.2.6.2. Identification of linkage by construction of IVM resistant Unc strains.....	98
3.2.6.3. Construction of doubly-marked strains for three factor mapping.....	98
3.2.6.4. Three factor mapping of IVM resistance mutations.....	99
3.2.6.5. Interpretation of three-factor mapping data.....	99
3.2.7. Complementation testing of nr272, nr2344, nr2477 and nr2389 against each other and other Dyf loci for the Dyf phenotype.....	100
3.2.7.1. Staining of worms and quantification of penetrance of the Dyf phenotype.....	100
3.2.7.2. Construction of double heterozygotes for DiO staining.....	100
3.2.7.3. Interpretation of complementation testing.....	101
3.2.8. Growth of dominant IVM resistance strains at 27°C.....	101
3.3. RESULTS.....	102
3.3.1. Mutagenesis screen for Dominant IVM resistance mutations.....	102
3.3.2. Lethal IVM dose for heterozygotes and homozygotes.....	103
3.3.3. Dyf phenotype of the dominant ivermectin resistant Dyf alleles nr272, nr2344, nr2477 and nr2389 as homozygotes and heterozygotes.....	105
3.3.3.1. Dyf phenotype	105
3.3.3.2. Original Strains (Appendix B).....	106
3.3.3.3. Outcrossed strains (Table 3.2.).....	106
3.3.3.4. Intensity of staining.....	107
3.3.4. Genetic mapping and complementation testing of dominant ivermectin resistance mutations.....	109

3.3.5. Growth of dominant IVM resistance strains at 27°C.....	114
3.4. DISCUSSION	116
3.4.1. Mutations conferring resistance to 5 ng/mL IVM in heterozygotes are rare.....	116
3.4.2. nr272, nr2389 and nr2477 heterozygotes are less resistant to IVM than homozygotes.....	116
3.4.3. Mutants isolated for dominant IVM resistance are Dyf, but the Dyf phenotype is incompletely penetrant in nr272, nr2344 and nr2477 heterozygotes, and Dyf is recessive for nr2389.....	118
3.4.4. Dominant IVM resistance mutations define two genetic loci, <i>dyf-10</i> I and <i>dyf-12</i> X.....	119
3.4.5. Male nr2389/+ heterozygotes are less resistant to IVM.....	121
3.4.6. Intergenic noncomplementation between <i>dyf-10</i> and <i>dyf-12</i>	122
3.4.7. IVM resistance of dominant mutations is temperature sensitive and also is associated with allele specific temperature effects on hermaphrodite viability.....	124
CHAPTER FOUR.....	126
4.1. INTRODUCTION	127
4.2. METHODS.....	133
4.2.1. Mutagenesis screen for suppressors of the Avr phenotype of <i>dyf-12(nr2477)d</i>	133
4.2.1.1. Experiment 1.....	133
4.2.1.2. Experiment 2.....	133
4.2.1.3. Re-screening of putative suppressed strains.....	134
4.2.1.4. Phenotypic Characterisation of Suppressor of Avr (<i>sav</i>) mutants.....	134
4.2.1.5. Mode of inheritance of <i>sav</i> mutants.....	134
4.2.1.6. Complementation testing of au7 with X-linked <i>dpy</i> mutations	135
4.2.1.7. Linkage analysis of <i>sav</i> mutations	135
4.2.2. Effect of <i>dpy</i> mutations on the Avr phenotype of Dyf strains.....	139
4.2.2.1. Construction of <i>dpy/dyf</i> double homozygotes.....	139
4.2.2.2. Dose response of <i>dpy/dyf</i> strains to IVM.....	141
4.2.3. Effect of <i>unc</i> mutations on the Avr phenotype of Dyf strains.....	141
4.3. RESULTS.....	142
4.3.1. Screen for suppressors of the Avr phenotype of <i>dyf-12(nr2477)</i>	142
4.3.2. Phenotypic Characterisation of <i>sav</i> (Suppressor of Avermectin resistance) mutants.....	143
4.3.3. Genetic Characterisation of <i>sav</i> mutations.....	145
4.3.4. Effect of <i>dpy</i> mutations on the Avr phenotype of Dyf strains.....	150

4.3.5. Effect of <i>unc</i> mutations on the Avr phenotype of Dyf strains.....	150
4.3.5.1. Kinesin gene mutations.....	150
4.3.5.2. Other <i>unc</i> mutations.....	152
4.4. DISCUSSION.....	154
4.4.1. <i>sav</i> mutations are common.....	154
4.4.2. <i>sav</i> mutants suppress IVM resistance of Dyf mutations but not the associated Dyf phenotype.....	154
4.4.3. nonUnc nonDpy <i>sav</i> mutations may interact with <i>unc</i> mutations.....	156
4.4.4. <i>dpy</i> mutations can suppress IVM resistance.....	156
4.4.5. <i>unc</i> mutations suppressing IVM resistance of Dyf mutants may be useful in elucidating the mechanism of resistance to IVM.....	158
4.4.5.1. Kinesin-like <i>unc</i> mutations.....	158
4.4.5.2. <i>unc</i> mutations involved in muscle structure.....	160
CHAPTER FIVE	162
5.1. Dominant ivermectin resistant mutations.....	163
5.2. A model for ivermectin resistance in Dyf mutants.....	165
5.2.1. PROPOSAL.....	165
5.2.2. EVIDENCE FOR THE MODEL.....	166
5.2.2.1. The most important site of IVM action for <i>C.</i> <i>elegans</i> in laboratory culture is probably the pharynx.....	166
5.2.2.2. Amphid defects in <i>C. elegans</i> confer resistance to low levels of IVM included in the growth media.....	169
5.2.2.3. In wild type worms the amphid may participate in pharyngeal regulation, but the pharynx is also self regulatory.....	172
5.2.2.4. The production of a constitutive signal from the amphid may be involved in IVM resistance mediated by Dyf mutations.....	176
5.2.5. EXPERIMENTS TO BE DONE.....	179
BIBLIOGRAPHY	184
APPENDICES	204
APPENDIX A.....	205
APPENDIX B.....	215

LIST OF FIGURES.

Figure 1.1.....	6
Figure 1.2. - Anterior arrangement of sensory neurons in <i>C. elegans</i> (White et al., 1986).....	21
Figure 1.3. - Appearance of amphid (Perkins et al., 1986), inner labial, outer labial and cephalic sensilla (Ward et al., 1975) from <i>C. elegans</i>	27
Figure 1.4.....	37
Figure 2.2. - Amphid neuron staining in a <i>H. contortus</i> L1 larva.....	63
Figure 2.3.....	64
Figure 2.4.....	69
Figure 3.1.....	108
Figure 4.1. Linkage analysis of <i>sav</i> mutations.....	137
Figure 5.1.....	167
Figure 5.2.....	173

LIST OF TABLES.

Table 1.2. - Some chemosensory mutants of <i>C. elegans</i>	24
Table 1.1. Neurons with dendritic endings in <i>C. elegans</i> amphids.....	30
Table 2.1. - Mutations in <i>unc</i> genes and other neuronal defective mutants tested for IVM resistance at three concentrations of the drug.....	53
Table 2.2. - Mutations which cause animals to have unusual levels of certain neurotransmitters and which were tested for IVM resistance at three concentrations.....	55
Table 2.3. Dye filling defective mutants, other chemosensory mutants and dauer defective mutants of <i>C. elegans</i> and their sensitivity to three concentrations of IVM.....	58
Table 2.4. Mutants of <i>C. elegans</i> which are Supersensitive to IVM.....	60
Table 2.5. Head Morphology mutants and their sensitivity to various concentrations of IVM.....	61
Table 2.6. - Amphid neuron staining patterns in L1 larvae of IVM resistant (CAVR) and susceptible (VRSG) strains of <i>Haemonchus contortus</i>	63
Table 2.7. Temperature sensitivity of ivermectin resistance in <i>unc-44</i> and nonUncdyf mutants.....	67
Table - 3.1. Dose response of outcrossed homozygotes and heterozygotes of four dominant IVM resistance mutations.....	104
Table - 3.2. Dyf phenotype of the dominant ivermectin resistant Dyf alleles nr272, nr2344, nr2477 and nr2389 as homozygotes and heterozygotes (outcrossed strains).....	107
Table - 3.3. Complementation tests (for the Dyf phenotype) of the dominant ivermectin resistant Dyf alleles nr272, nr2344 and nr2477 against alleles of previously defined Dyf loci.....	111
Table - 3.4. Complementation tests (for the Dyf phenotype) of the dominant ivermectin resistant Dyf allele nr2389 against alleles of previously defined Dyf loci.....	112
Table - 3.5. Three Factor Genetic Mapping of the dominant IVM resistant alleles nr272, nr2344, nr2477 and nr2389.....	113

Table 3.6. Temperature sensitive sterility and temperature sensitive IVM resistance of dominant IVM resistant mutants.....	115
Table 4.1. Results of two ems mediated mutagenesis screens and subsequent rounds of screening for mutants suppressing ivermectin resistance conferred by <i>dyf-12(nr2477)</i>	144
Table 4.2. Summary of mutant strains generated in the two ems-mutagenesis experiments screening for suppressors of the Avr phenotype of <i>dyf-12(nr2477)</i>	149
Table 4.3. - Suppression of the IVM resistance phenotype of dominant alleles of <i>dyf-10</i> and <i>dyf-12</i> by <i>dpy-7</i> and <i>dpy-8</i>	151
Table 4.4. - Suppression of the IVM resistance phenotype of Dyf alleles by <i>unc</i> mutations.....	153

A BSTRACT.

Ivermectin (IVM) is a potent anthelmintic which is lethal to *Caenorhabditis elegans* when included in nutrient agar growth medium at concentrations above 2 ng/mL. Recessive mutations imparting resistance to 5 ng/mL are obtainable in "1 out of 204 mutagenised genomes" (0.05 M ethyl methanesulfonate) and arise spontaneously at 1.7×10^{-5} (Kim and Johnson, 1991). These "low-level" resistance mutations (resistant to 5-30 ng/mL IVM) occur at about thirty loci (Kim and Johnson, 1991). Dominant IVM resistant alleles have been observed under field conditions in *Haemonchus contortus* and these are resistant to low levels of IVM in *in vitro* larval development assays (Lacey, Redwin *et al.*, 1990; Martin and Turney, 1992; LeJambre, 1993; LeJambre, Gill *et al.*, in press). *C. elegans* was used as a model to study IVM resistance in parasitic nematodes. To identify loci at which dominant IVM resistance alleles occur, a *C. elegans* mutagenesis screen was undertaken to obtain dominant IVM resistance (5 ng/mL) alleles. The rate of mutagenesis for dominant IVM resistance was found to be 7.9×10^{-7} , indicating that these alleles are much rarer than recessive alleles.

Characterisation of the dominant IVM resistance alleles revealed that they have varying levels of IVM resistance as homozygotes and heterozygotes. Three of four alleles isolated (nr272, nr2389 & nr2477) are incompletely dominant for IVM resistance and the IVM lethal dose for heterozygotes and homozygotes of the fourth allele (nr2344) were identical. IVM resistance of heterozygotes was greatest when the resistance allele was inherited paternally for nr272 and nr2389 and heterozygote resistance was greatest when the resistance allele was inherited maternally for nr2477. Finally, IVM resistance is observed in nr2389/+ hermaphrodites but not in nr2389/+ males (*i.e.* the dominance is sex limited).

The four dominant IVM resistant alleles have a Dyf (dye filling defective) phenotype as homozygotes, indicating that the mutations may cause structural or functional abnormalities in the amphids. The Dyf phenotype is recessive for nr2389 and incompletely dominant for nr272, nr2344 and nr2477; the number of Dyf heterozygotes is low and the staining in the remainder is fainter than for wild type worms. Subsequent to these findings a large number of previously characterised mutant strains were analysed for IVM resistance, and it was found that mutations at 27 loci which are Dyf are also resistant to 5 ng/mL IVM. Alleles of three *unc* loci also are IVM resistant and have faint amphid staining. nr2389 fails to complement the Dyf phenotype of *dyf-10*(e1387) and nr272, nr2344 and nr2477 fail to complement the Dyf phenotype of one another and *dyf-12*(sa127). Three factor interval mapping using the IVM resistance phenotype placed nr2389 on linkage group I near *dyf-10* and nr272, nr2344 and nr2477 on linkage group X near *dyf-12*. The complementation and mapping data support nr2389 being an allele of *dyf-10* and nr272, nr2344 and nr2477 being alleles of *dyf-12*. The dominant IVM resistant alleles of *dyf-10* and *dyf-12* isolated here fail to complement one another for the Dyf phenotype, which suggests an interaction between the two genes in amphid neuron formation or function.

Some mutations which impart resistance to very low levels of IVM (between 2 and 5 ng/mL) have also been identified, these mutations have Tax, Bli, Notch or Rounded nose phenotypes and may result in slight alterations to amphid morphology or function. A group of mutations which are supersensitive to IVM were also identified, most notable among these were alleles of six *eat* loci. The super sensitivity of Eat mutants, along with the findings of a number of other studies implicate the pharynx as a major site of action of IVM. *unc-104*(e1265) suppresses IVM resistance (but not Dyf) in a range of Dyf strains (this study) and *unc-116*(e2310), *snt-1*(md290) and *snt-1*(md325) also suppress IVM resistance (but not Dyf) in some Dyf strains (Grant Pers. Comm., 1995). These mutations which suppress IVM resistance in Dyf worms are involved in synapse function, suggesting that proper synapse function is necessary for IVM resistance.

Information provided by the experiments described here and from published studies of the mode of action of IVM, suggest a mechanism for IVM resistance in *dyf* mutants of *C. elegans*. I propose that IVM resistance in *C. elegans* is dependent on a

constitutive neuronal (or humoral) signal from the amphids which stimulates the pharynx decreasing the pharyngeal response to IVM. This hypothesis can be readily tested in the laboratory, and a number of experiments are discussed which might reveal more about amphid-mediated control of the pharynx and IVM resistance in *Dyf* worms.

AIMS

1. To isolate and characterise mutants carrying dominant ivermectin resistant mutations in *Caenorhabditis elegans* comparable to dominant IVM resistant mutations in *Haemonchus contortus*.
 2. To investigate the nature of ivermectin resistance in *Caenorhabditis elegans*.
-