The integration of the parasitoid *Microplitis demolitor* Wilkinson (Hymenoptera: Braconidae) into integrated pest management systems in Australian cotton

Robert Andrew Annetts B.Sc. (Honours) The University Of New England

October 2000

A thesis submitted for the degree of Doctor of Philosophy of the University of New England

Declaration

I certify that the contents of this thesis have not been submitted for any other degree and are not currently submitted for any other degree.

I certify that all sources of information used have been acknowledged in this thesis.



Robert Annetts.

Acknowledgments

Very special thanks to my supervisor Dr. David Murray, who provided guidance, advice, encouragement and whose knowledge and patience is unquestionable. Thanks also to my University based supervisor associate Professor Peter Gregg who provided excellent advice and ideas, especially when correcting the final manuscript. Thanks to Natalie Martin for reading and correcting early drafts, and David Annetts and Matt Cahill for correcting later drafts and providing valuable criticism.

It is impossible to name everyone who has contributed in some way to this work. So if I've forgotten anyone who has contributed, sorry. I would especially like to thank Kristen Latimer for providing expert assistance in the field during 1996/97. Special thanks also to Sue Maclean, Brad Scholz and Richard Lloyd for technical assistance, support and encouragement throughout my term. I would also like to thank Andrea Gerrard, Angela Doherty and Kerry Rynne for technical assistance, Simon Hoyle and Kerry Bell for statistical analysis and Ian Newton for help with ascovirus literature and sharing ideas on this topic.

I would like to thank Dow AgroSciences and Rohm and Haas for providing technical grade spinosad and methoxyfenoxyzide insecticides respectively, when they were early in development. Also Crop Care, Bayer and NSW Agriculture (Neil Forrester) for providing other technical grade products. Thanks to Murray Boshammer (Total AGricultural Services, TAGS) for allowing me to use *Helicoverpa* spp. data and the Farming Systems Institute, Queensland Department of Primary Industry (QDPI), and John Marshall, then with the QDPI, Dalby, for contributing data.

The manual for CAPTURE (White *et al.*, 1982) and an associated publication (Otis *et al.*, 1978) were kindly supplied by G. White. Thanks also to G. White for analysing the capture/recapture data.

Thanks on a personal note to my parents for support and encouragement, and to Raisa Silvonen for support, encouragement and putting up with me. Thanks also to Dr. Mary Notosteine, my honours supervisor, who really encouraged me to do a Ph.D. in the first place.

Thanks to Jeff and Marilyn Bidstrup for cooperation and support at the Warra trial site and Bill Arthur for cooperation and support at the Nandi site. I would like to thank the CRC for Sustainable Cotton Production, who funded this project (DPIQ223).

Table of contents

TRODUCTION 1

CHAPTER 1

Cotton Production in Australia	4
Introduction	
What is cotton?	
Why is cotton cultivated?	
Where is cotton grown in Australia?	
How is cotton grown in Australia?	5
Conclusions	6

CHAPTER 2

Helicoverpa: A review	7
Introduction	7
Why are Helicoverpa spp. such devastating pests?	7
Identification	
Distribution	
Crops attacked	
Feeding habits	
Fecundity and development	9
Diapause	9
Mobility	
Resistance in Helicoverpa spp	
Resistance management strategy	
Economic impact of Helicoverpa spp	
Future	
Conclusions	

Integrated Pest Management of Helicoverpa spp. in Australian cotton	13
Introduction	
Insecticides in IPM	14
Chemical control of Helicoverpa spp	14
Side-effects of insecticide use targeted at Helicoverpa spp	
Cultural control	

Crop management	
Other methods of cultural control	
Host plant resistance	
Genetically modified or Ingard [®] cotton	
Compensation	
Natural mortality	
Biological control of Helicoverpa spp	
Pathogens of Helicoverpa spp.	
Natural enemies of Helicoverpa spp	
Classical biological control	
Augmentation	
Naturally occurring heneficials	21
Predators	
Predators Parasitoids	23
Predators Parasitoids M. demolitor	23
Predators Parasitoids M. demolitor Managing beneficials	23

M. demolitor: A key component of IPM in Australian cotton	
Introduction	
Biology and ecology of M. demolitor	
Host range	
Life-cycle	
Sex determination	
Mate location	
Host location	
Superparasitism	
Effects on the host	
Ascoviruses	
Diurnal Behavior	
Estimating populations in the field	
Monitoring in the field	
Percent parasitism	
Effects of pesticides	
Conclusions	

Food consumption and weight gain of *H. armigera* larvae after parasitisation by *M*.

demolitor	
Abstract	
Introduction	
Materials and Methods	
Insects	
Trial	
Results	
Discussion	
Conclusions	

Monitoring M. demolitor in the field	
Abstract	
Introduction	
Methods of monitoring parasitoids in the field	
Attractive colours	
Suction machines	
Percent parasitism of the host	45
Pheromones	45
Direct observations	
Materials and Methods	
Field sites	
Farming Systems Institute, QDPI, IPM trial site (Appendix 4)	
Biological data	
Plant measurements	
Helicoverpa spp. egg and larval counts	
Spray histories	
Sampling methods	
Suction samples	
Yellow coloured traps	
Traps based on host insects	
Sticky traps baited with virgin female M. demolitor	
Percent parasitism	
Direct observations	
Results	
Biological data	
Plant data	
Helicoverpa spp. egg and larvae counts	

Spray history	60
Sampling methods	60
Suction samples, yellow coloured water traps and sticky traps baited with H. armigera larvae	60
Sticky traps baited with virgin females	
Fate of collected larvae and percent parasitism	
Direct observations	
Discussion	79
Sticky traps baited with virgin females	80
Percent parasitism	82
Direct observations	84
Conclusions	85

Preliminary estimation of adult populations of *M. demolitor* in an unsprayed cotton field 86

Abstract	86
Introduction	86
Materials and methods	87
Results	87
Discussion	90
Conclusions	93

CHAPTER 8

Diurnal activity of <i>M. demolitor</i> adults in the cotton crop	95
Abstract	
Introduction	
Materials and methods	
Sampling dates	
Sampling methods	
Direct observations (sweep netting)	96
Sticky traps baited with virgin female M. demolitor	
Results	
Discussion	101
Conclusions	

Preliminary study of the release-recapture of male <i>M. demolitor</i> adults	
Abstract	104
Introduction	104
Materials and methods	105

Sampling methods	105
Results and Discussion	106
Conclusions	110

Base-line susceptibility and calibration of discriminating doses for *H. armigera* larvae to

selected insecticides	
Abstract	
Introduction	
Materials and methods	
Insects	
Bioassay	
Stomach poisons	
Topical testing	
Analysis	
Sub-lethal effects	
Results	
Bioassay	
Stomach insecticides	
Contact insecticides	
Sub-lethal effects after exposure to insecticides	
Discussion	
Bioassay	
Sub-lethal effects	
Conclusions	

Toxicity of insecticides to adult and pupal <i>M. demolitor</i>	
Abstract	
Introduction	
Materials and Methods	137
Insects	
Bioassay	
Results	139
Adult stage	
Wasp weights	
Pupal stage	
Discussion	

Conclusions1	149
--------------	-----

Abstract	
Introduction	
Materials and methods	
Insects	
Bioassay	
Topical testing	
Stomach poisons	
Analysis	
Results	
Discussion	
Conclusions	
NCLUSIONS	

APPENDIX 1

Insect rearing	
M. demolitor	195
H. armigera and C. argentifera	
Diet preparation	
Standard Heliothis diet	
Mould inhibitor preparation	
Sugar solution for moths	
References	

APPENDIX 2

Insecticides		
Topical testin	1g	
Stomach pois	sons	

APPENDIX 3

Equipment register	200
---------------------------	-----

APPENDIX 4

1 1010 51005	
1994/95	202
Warra site	202
Yield and brief review	203
Nandi site	203
Yield and brief review	204
1995/96	205
Warra	205
Yield and brief review	206
Dalby	206
1996/1997	207
Warra	207
Yield and brief review	209

APPENDIX 5

List of Figures

Figure 4-1. Life cycle of <i>M. demolitor</i> (drawn by Dr. D. Murray).	30
Figure 4-2. Diagrammatic representation distinguishing the sexes of M. demolitor. Abdomen of ma	le and female M.
demolitor, showing ovipositor protruding in the female.	32
Figure 5-1. Diet consumption by second instar (at day 0) H. armigera larvae unparasitised and para	isitised by M.
demolitor (error bars are standard errors of the means)	42
Figure 5-2. H. armigera larval weights, larvae unparasitised and parasitised by M. demolitor (error	bars are standard
errors of the mean).	43
Figure 5-3. Numbers of M. demolitor larvae emerging from H. armigera hosts (days after parasitisa	ution)43
Figure 6-1. Cotton development calendar from a dryland crop at Warra, southeast Queensland during	ng 1994/95 (Data
courtesy of Dr. D. Murray, Farming Systems Institute, QDPI). Boxed area is the critical period	l for insect
control	56
Figure 6-2. Number of Helicoverpa spp. per metre in the unsprayed block at Warra during 1994/95	(data courtesy of
M. Boshammer, Total AGricultural Services (TAGS)	57
Figure 6-3. Number of Helicoverpa spp. per metre at the unsprayed block at Nandi during1994/95	(data courtesy of
QDPI, Farming Systems Institute and John Marshall, QDPI, Dalby)	57
Figure 6-4. Number of Helicoverpa spp. per metre at the unsprayed block at Warra during 1995/96	(data courtesy of
TAGS).	58

Figure 6-5. Number of Helicoverpa spp. per metre at the unsprayed block at Warra during 1996/97 (data courtesy of Figure 6-6. Number of Helicoverpa spp. per metre at the refuge block at Warra during 1996/97 (data courtesy of Figure 6-7. Number of Helicoverpa spp. per metre at the refuge block at Witu during 1996/97 (data courtesy of Figure 6-8, Number of male *M. demolitor* caught in sticky traps, baited with virgin females. Warra 1994/95, results Figure 6-9. Number of male *M. demolitor* caught in sticky traps, baited with virgin females, at Nandi during 1994/95. See Table 6-8 for number of traps on each sampling occasion. Numbers refer to application of possible Figure 6-10. Number of male M. demolitor caught in sticky traps baited with virgin females from Warra IPM trial site during 1995/96. See Table 6-5 for number of traps on each sampling occasion. Numbers refer to Figure 6-11. Number of male *M. demolitor* caught in sticky traps baited with virgin females at the Warra IPM site and Warra refuge site during 1996/97. See Table 6-7 for number of traps on each sampling occasion. Numbers refer to application of possible disruptive insecticides, see Table 6-4......70 Figure 6-12. Number of male *M. demolitor* caught in sticky traps baited with virgin female at the Witu refuge site during 1996/97. See Table 6-7 for number of traps on each sampling occasion......70 Figure 6-13. Fate of Helicoverpa spp. larvae collected from the unsprayed block at Warra during 1994/95.....72 Figure 6-14. Fate of collected Helicoverpa spp. larvae collected at Nandi during 1994/95......72 Figure 6-16. Fate of collected Helicoverpa spp. larvae at the unsprayed block at the Warra IPM trial site, during Figure 6-17. Fate of *Helicoverpa* spp. larvae collected at unsprayed block at Nandi during 1995/96......74 Figure 6-18. Fate of Helicoverpa spp. larvae collected from the unsprayed block at the Warra IPM trial site, during Figure 6-19. Fate of Helicoverpa spp. larvae collected at the Warra refuge block during 1996/97......76 Figure 6-20. Fate of *Helicoverpa* spp. larvae collected at the Witu refuge site during 1996/97......77 Figure 6-21. Number of M. demolitor caught in sweep net per 30 minutes (100m) searching at the unsprayed block at the Warra IPM trial site during 1996/97......78 Figure 6-22. Number of M. demolitor caught in sweep net per 30 minutes (100m) searching at the Warra refuge site during 1996/97......78 Figure 6-23. Number of M. demolitor caught in sweep net per 30 minutes (100m) of searching at the Witu refuge site during 1996/97......79 Figure 7-1. Number of male and female M. demolitor adults caught in sweep netting and percent parasitism of collected Helicoverpa spp. larvae from unsprayed refuge block at Warra during 1996/97. These data were presented in Chapter 6......90

Figure 8-1. Diurnal activity of male and female M. demolitor in a conventionally sprayed block at Warra on the 31 Figure 8-2. Diurnal activity of male and female *M. demolitor* in and unsprayed block at Dalby on the 19 March 96 Figure 8-3. Diurnal activity of male and female *M. demolitor* in an unsprayed block at Warra refuge on the 15 Figure 8-4. Diurnal activity of male and female *M. demolitor* in unsprayed refuge block at Witu on 4 April 97 (1 Figure 8-5. Diurnal activity of male and female *M. demolitor*, average of all studies......100 Figure 8-6. Average number of male M. demolitor caught per trap per hour from 0600 to 0600 the following day in an unsprayed irrigated block at Dalby (19 March 96). Sampling was carried out hourly from 0600 to 1800, and traps were sampled at 0600 the following day.....100 Figure 8-7. Average number of male *M. demolitor* caught per trap per day (19 to 21 March 96) in an unsprayed irrigated block at Dalby (Number of traps on each sampling occasion is displayed in each graph).........101 Figure 9-1. Arrangement of sticky traps baited with virgin females (view from above). R= release point......106 Figure 9-2. Arrangement of mark-recapture grid relative to the normal traps showing the number of wasps caught in Figure 9-3. Total number for each axis of the grid of male M. demolitor caught in sticky traps baited with virgin Figure 10-1. Weight and mortality of H. armigera larvae, after dosing with B. thuringiensis. Error bars for larval weight are standard errors of the means and error bars for percent mortality are 95% confidence limits. 123 Figure 10-2. Weight and mortality of H. armigera larvae, after dosing with chlorfluazuron. Error bars for larval weight are standard errors of the means and error bars for percent mortality are 95% confidence limits. 123 Figure 10-3. Weight and mortality of H. armigera larvae, after dosing with lufenuron. Error bars for larval weight Figure 10-4. Weight and mortality of H. armigera larvae, after dosing with spinosad (NAF-85). Error bars for larval weight are standard errors of the means and error bars for percent mortality are 95% confidence limits. 124 Figure 10-5. Weight and mortality of H. armigera larvae, after dosing with spinosad (XDE-105). Error bars for larval weight are standard errors of the means and error bars for percent mortality are 95% confidence limits. Figure 10-6. Weight and mortality of H. armigera larvae, after dosing with methoxyfenozide. Error bars for larval weight are standard errors of the means and error bars for percent mortality are 95% confidence limits. 125 Figure 10-7. Weight and mortality of H. armigera larvae, after dosing with profenofos. Error bars for larval weight Figure 10-8. Weight and mortality of H. armigera larvae, after dosing with thiodicarb. Error bars for larval weight

Figure 10-9. Days after dosing with sub-lethal concentrations of chlorfluazuron until pupation of secon	d instar H.
armigera larvae	127
Figure Appendix 4-1. Layout and treatment sizes for the Warra IPM trial site, 1994/95	203
Figure Appendix 4-2. Layout and plot sizes for trial site at Nandi, 1994/95	204
Figure Appendix 4-3. Layout and plot sizes of the Warra IPM trial site, 1995/96.	206

List of Tables

Table 6-1. Number of <i>M. demolitor</i> caught in suction samples (500 metres), at Warra IPM trial site, during the
1994/95 season61
Table 6-2. Number of <i>M. demolitor</i> caught in yellow coloured water traps in the unsprayed block at Warra IPM trial
site, during the 1994/95 season62
Table 6-3. Number of M. demolitor caught in traps baited with live H. armigera larvae in unsprayed Ingard® refuge
blocks, during the 1995/96 season62
Table 6-4. Number of sticky traps at Warra and Nandi during 1994/95. 1 female/trap unless noted, checked after 7
days65
Table 6-5. Number of sticky traps at the IPM site at Warra during 1995/96. 1 female/trap, checked after a day.67
Table 6-7. Average number of M. demolitor caught in sticky traps baited with virgin females at Dalby during
1995/96
Table 6-7. Number of sticky traps baited with virgin females at the Warra IPM site, Warra refuge site and Witu
refuge site during the 1996/97. 1 female per trap, checked after 1 day
Table 6-8. Number of larvae collected on each sampling occasion at Warra and Nandi during 1994/9571
Table 6-10. Number of Helicoverpa spp. larvae collected at the Warra IPM trial site and Nandi during 1995/96.
Table 6-12. Number of Helicoverpa spp. larvae collected from the unsprayed block at the Warra IPM trial site,
Table 6-12. Number of Helicoverpa spp. larvae collected from the unsprayed block at the Warra IPM trial site, Warra refuge and Witu refuge during 1996/97
Table 6-12. Number of Helicoverpa spp. larvae collected from the unsprayed block at the Warra IPM trial site, Warra refuge and Witu refuge during 1996/97
 Table 6-12. Number of <i>Helicoverpa</i> spp. larvae collected from the unsprayed block at the Warra IPM trial site, Warra refuge and Witu refuge during 1996/97
 Table 6-12. Number of <i>Helicoverpa</i> spp. larvae collected from the unsprayed block at the Warra IPM trial site, Warra refuge and Witu refuge during 1996/97
 Table 6-12. Number of <i>Helicoverpa</i> spp. larvae collected from the unsprayed block at the Warra IPM trial site, Warra refuge and Witu refuge during 1996/97
 Table 6-12. Number of <i>Helicoverpa</i> spp. larvae collected from the unsprayed block at the Warra IPM trial site, Warra refuge and Witu refuge during 1996/97
 Table 6-12. Number of <i>Helicoverpa</i> spp. larvae collected from the unsprayed block at the Warra IPM trial site, Warra refuge and Witu refuge during 1996/97
 Table 6-12. Number of <i>Helicoverpa</i> spp. larvae collected from the unsprayed block at the Warra IPM trial site, Warra refuge and Witu refuge during 1996/97
 Table 6-12. Number of <i>Helicoverpa</i> spp. larvae collected from the unsprayed block at the Warra IPM trial site, Warra refuge and Witu refuge during 1996/97
 Table 6-12. Number of <i>Helicoverpa</i> spp. larvae collected from the unsprayed block at the Warra IPM trial site, Warra refuge and Witu refuge during 1996/97

Table 11-2. Mortality of <i>M. demolitor</i> pupae (%) dosed with acetone, one, 10, and 100 times the LD_{50} determined
for M. demolitor adults (see Table 11-1)
Table 12-1. Egression of <i>M. demolitor</i> (%) from parasitised <i>H. armigera</i> larvae and host mortality (%) from
unparasitised H. armigera larvae (number treated) treated with the LC ₉₉ (see Chapter 10, Table 10-1) of
selected stomach insecticides161
Table 12-2. Egression of M. demolitor from parasitised H. armigera larvae or host mortality (%) from unparasitised
H. armigera larvae treated with the LD ₉₉ (see Chapter 10, Table 10-2) of selected contact insecticides162
Table Appendix 3-1. Equipment used in this thesis. 200
Table Appendix 5-1. Spray history of the IPM site at Warra during 1994/95. Trade name (active ingredient).210

Table Appendix 5-2. Spray history for Nandi during 1994/95. Trade name (active ingredient)......211

Table Appendix 5-3. Spray history for the IPM site at Warra during 1995/96. Trade name (active ingredient).212

Table Appendix 5-4. Spray history for the IPM site at Warra during 1996/97. Trade name (active ingredient).213

Abstract

Cotton production in Australia, the ecology of the key pests of Australian cotton, *Helicoverpa* spp., and integrated pest management (IPM) in Australian cotton were reviewed. *Microplitis demolitor* Wilkinson (Hymenoptera: Braconidae) is a key beneficial and a critical component for the success of IPM in Australian cotton. The ecology of *M. demolitor* and effects of insecticides on the life stages of *M. demolitor* were studied, in an attempt to promote conservation and increase the role of *M. demolitor* in IPM.

This study showed that *Helicoverpa* spp. larvae parasitised by *M. demolitor* cause insignificant damage compared to unparasitised larvae, with a parasitised larva consuming approximately 5% of that consumed by a healthy larva. This means that parasitised larvae should be tolerated and not counted in larvae checks to determine spray decisions. *M. demolitor* larvae took 10 days to complete larval development at 25°C, 60-70% relative humidity and a 14:10 light: dark photoperiod. These data contribute to the basic understanding of the ecology of *M. demolitor*.

M. demolitor was the dominant larval parasitoid in southeast Queensland and was present in the cotton crop at the critical stage of the crop's development. *M. demolitor* appeared in the crop early November to December, and occurred in significant numbers in the crop from early to mid December until the end of the season. High rates of parasitism of *Helicoverpa* spp. (between 50% and 90%) of second to third instar *Helicoverpa* spp. larvae by *M. demolitor* were recorded. Relatively large numbers of *M. demolitor* adults were found in sprayed fields compared to unsprayed fields.

Methods of monitoring *M. demolitor* adults and the impact of *M. demolitor* on *Helicoverpa* spp. larvae in the field were studied. It was determined that an estimate of percent parasitism, sticky traps baited with virgin females and direct observations were useful tools for research or for ongoing monitoring of *M. demolitor*. Suction sampling, yellow coloured water traps and traps baited with *H. armigera* larvae proved unsuccessful at monitoring *M. demolitor*.

The diurnal behaviour of M. demolitor adults was studied. Large numbers of adult M. demolitor were observed in the field. Both male and female M. demolitor were inactive early in the

morning and late evening, but were equally active throughout the day. There was a distinct diurnal pattern in catches from sticky traps baited with virgin females. Males were caught most often early in the morning, declining during the afternoon. None were captured during the night. Disruptive insecticides should be applied when *M. demolitor* are inactive. In a monitoring program, adult *M. demolitor* could be monitored at any time of the day without biasing results.

A mark-recapture study estimated the *M. demolitor* population in an unsprayed cotton field. The population of *M. demolitor* males present in the 5 ha. block of unsprayed cotton was estimated to be between 1503 and 2421 (ca. 300-484 wasps/ha.), and the female population was estimated at 365 by extrapolating from the ratio of male wasps to female wasps at the time of the study numbers. At the time of the study there was approximately 60% parasitism of *H. armigera* larvae. This study showed that a small number of female *M. demolitor* in the field contributed to high parasitism rates of the host. This indicates that inundative releases of female *M. demolitor* may be a economically feasible control option for *Helicoverpa* spp. The mark-recapture method used in this study is useful for investigating the effects of pesticide treatments, by allowing estimation of the population present before and after insecticide application.

A release-recapture study of male *M. demolitor* was carried out. This study showed that fluorescent powder could be successfully used in a release-recapture study of *M. demolitor*, and most likely any medium sized Hymenoptera. This study showed that catch efficiency of the sticky traps baited with virgin females was reduced after 2 days in the field. Male *M. demolitor* were most likely to be caught in the closest sticky trap baited with virgin female, with male *M. demolitor* males not moving very far in response to these traps. This study showed that there was a large natural population of male *M. demolitor* wasps (ca. 62.5 male *M. demolitor* adults/ha.) present in a field sprayed only with biologically active insecticides.

M. demolitor prefer to parasitise second and third instar *Helicoverpa* spp. larvae. In order to study the interactions between *M. demolitor* larvae, host larvae and insecticides, baseline dose-response data were generated on second instar *Helicoverpa* spp. larvae. The IGR compounds, chlorfluazuron and lufenuron, as well as the MAC compound, methoxyfenozide, and Naturalyte, spinosad were the most effective stomach (ingestion-active) compounds tested against *H.*

armigera. These data are a useful reference tool and were used in the study on the interactions between *M. demolitor* larvae, host larvae and insecticides. The results are significant for IPM. The sub-lethal data generated in this trial show that although affected larvae do not die, their weights are significantly reduced, demonstrating reduced damage in the crop. This means that larvae affected by sub-lethal doses will cause negligible damage in the crop, and may be suitable sources of food for predators or hosts for parasitoids. The implications for insecticide resistance management are discussed.

A study examining the affects of commonly used cotton insecticides on *M. demolitor* throughout its life cycle was carried out. It was found that adult *M. demolitor* is tolerant of some insecticides. The toxicities of tested compounds for adult *M. demolitor*, in ascending order, were: chlorfluazuron = methoxyfenozide < primicarb < endosulfan \leq dimethoate \leq cyhalothrin < profenofos < deltamethrin \leq bifenthrin < spinosad. This study showed that *M. demolitor* was relatively more tolerance of certain insecticides during its larval and pupal stages. *M. demolitor* pupae were protected from the effects of insecticides, except the pyrethroids. This study showed *M. demolitor* larvae developing within parasitised host larvae were unaffected by insecticide applications. However, larval parasitoids were indirectly affected by insecticides through host mortality. Larval parasitoids did not increase the susceptibility of their host to the insecticides, and parasitisation actually reduced the susceptibility of the host larvae to stomach insecticides, and to a lesser degree to contact insecticides.

This study showed that only the IGR compounds and the carbamate, pirimicarb, were of lower toxicity to the parasitoid than the larval pest. None of the other insecticides tested in the present study were acceptable for use in an IPM program. However, targeting insecticide applications when *M. demolitor* is in the pupal or larval stage will reduce the impact of most disruptive insecticides. The use of insecticides with slow rates of kill may promote parasitism by allowing parasitoid larvae enough time to complete development.