Using turbidity tolerant water plants in the rehabilitation of turbid farm dams

Annabel Douglas-Hill Graduate Diploma in Natural Resources University of New England

A thesis submitted for the degree of Master of Resource Science at the University of New England

March 2003

Frontispiece

Son: I'm sick of the Internet. I want a yabby net.

Father: Well you can't have one. It would corrupt you. You would end up staring into some strange dark pond all day: some pond full of reeds and mud......and mysterious life forms.

Then you would stare at the reflection on the water and see the sky, the clouds and the birds all quite differently.

You would throw a stone into the pond causing the reflection to ripple and distort. And gazing at it you would fall into a trance of wonderment and delight and never fit into normal life again.

Sorry - no yabby net!

Michael Leunig

Declaration

To the best of my knowledge and belief, this thesis contains no material previously submitted for a degree or any other award, in any university by any person, or any material previously published or written by any other person, except where due reference is made in the text. I consent to the thesis being made available for copying and loan if accepted for the award of the degree provided due acknowledgement is given.

Annabel Douglas-Hill

March 2003

Acknowledgments

This thesis is the result of financial support over the two years of the research from the Land and Water Resources Research and Development Program and from the Botany Department at the University of New England, through their Noel C.W. Beadle Scholarship.

I thank Anthony Casanova and Dr. Donald McLeod respectively for sharing their understanding of soils and clay colloids and for divulging to me the freely available sources of clay which was used in the pond experiments. I appreciate the communications with Dr. Stuart Blanch on the practical problems of keeping water turbid. I am indebted to many in the Botany and Ecosystem Management Departments: Chris Cooper for his technical skills in computers and water plant identification, Matt Dickinson for his help with treating the farm dams, Marion Costigan for her advice on phosphate assays, Doug Clark for his ingenious skills in solving all things mechanical and Olive Bourke for her personal concern over my health and safety when wading in farm dams. Many thanks to Katharine Crosslé and Geradine Woods for helping with the layout and formatting of this thesis and to the postgraduates Kerry, Michelle, Kathy and Kirsten for giving of their support and advice. I wish to thank Dr. Stuart Cairns of the Zoology Department for his guiding hand in the statistical interpretation of the farm dam assays, Rob Nethery of CSIRO for his tutoring in computer graphics, Laurie O'Donnell for his comments on my writing. Bronwyn Jones of the Department of Land and Water Conservation saved me from despair by finding computer programs that complemented mine.

Drs. Margaret Brock and Michelle Casanova undertook the principal role of my supervision and were tireless in sharing their wealth of knowledge, experience and enthusiasm on wetland plants and water issues and for their constructive comments on my writing. I thank Margaret for sourcing the funding for my research and for her faith in my ability to undertake it. Michelle taught me to identify the smallest water plants, including her beloved charophytes (stoneworts), and assisted with dam sampling and treatment. After my move to the Philippines, I greatly appreciate Michelle's patience and inspiration conveyed over the distance through the Internet. In their special way, Michelle and Margaret have corrupted me by giving me that yabby net.

Special thanks go to the Landcare farmers, Chris and Maria Eveleigh, David and Alex Taylor, Kim Barnet, Sonia and Ted Williams, Deanna Forster, Julie and David Kennelly, Alanna and Barb of Pooh's Nursery and Sue Burgess who gave me access to dams on their properties. Without them this research would not have been as worthwhile. Chris and Maria distributed information and cups of tea. David and Alex Taylor and Sonia and Ted

Williams organized the field day on water quality and aquatic ecology on their respective properties.

I thank my children Cassie and Ken, for their love and support when undertaking their own demanding studies and for their help with sampling, and along with my stepchildren, Rowan and Euan, for making the Christmas breaks good ones. Thanks for convincing me that I had to finish! Thanks to Cassie and Chelly for keeping me company in the sweltering monsoon nights during the writing-up stage. Aurea Dayaday and Victor Valentino, who diligently kept our home clean and running in my absences, have my grateful recognition.

I thank Priscilla Piper for chauffeuring me on a daily basis and for her positive attitude and Narelle Morse for editing my work despite her own pressing schedule. Michael and Molly Hillard adopted me as 'family' and gave freely of their friendship and emotional support through the last two-weeks-that-turned-into-two-months. This, plus meeting my daily needs for sustenance; the red wine, gossip and laughs has left me with many wonderful memories. It would take me a lifetime to thank you both enough.

Finally, I dedicate this thesis to my husband, Douglas Gray, for his depth of love, patience, support and encouragement and for his experience with tables and graphs. His daily emails sent from wherever he was based in Asia sustained my resolve and ensured that this thesis was completed.

Abstract

The colonization of some Australian wetlands by submerged water plants appears to be restricted by high water turbidity, which reduces underwater light and this turbidity appears to be a permanent state in many wetlands. A relationship between the lack of submerged water plant cover and high phytoplankton concentrations, particularly of blue-green algae, has been observed in turbid farm dams. Water plants are known to reduce algal blooms, clarify the water and improve water quality, but the processes of submerged water plant establishment are not fully understood.

This research project set out to investigate the influence of turbidity on submerged water plant germination and establishment and to find turbidity tolerant species that could be used to rehabilitate farm dams. Two seed banks, one from a natural but temporary wetland, and one from a permanent man-made wetland, were examined for water plant germination and establishment under four turbidity and three depth treatments in artificial ponds. The underwater Photosynthetically Active Radiation (PAR) was found to be reduced to below that needed for photosynthesis in about 30 cm of water with a mean turbidity of 135 nephelometric turbidity units (NTU). Germination from the two wetland seed banks showed no reduction in species richness with increasing turbidity under constant depth, but long periods of turbidity would probably affect the composition of the water plant community. Four submerged charophyte species (plant-like algae) were stimulated by an increase in turbidity to significantly increase their numbers, biomass and length while angiosperm species were suppressed by the permanent flooding regime. Charophytes (also known as stoneworts) from the seed bank of the temporary wetland developed the highest biomass, whereas in the seed bank from the permanent wetland, the submerged angiosperm, Vallisneria gigantea, had the greater biomass. The species that dominated germination from the seed bank in the pond and glasshouse trials could be predicted from observing the species in the wetland from which the seed bank was sourced and the water regime. This study suggests that charophytes may be adapted to turbid conditions. By using the functional classification method for wetland plants of Brock and Casanova (1997) it was shown that most 'fluctuation tolerators' (emergents and low growing species) germinated and established in the shallows where there was sufficient PAR. 'Fluctuation responders' (plastic and floating plants) however, were able to grow from a greater depth and emerge above the turbid water to photosynthesize and reproduce. Two submerged angiosperms, Vallisneria gigantea and Potamogeton ochreatus, were found to be tolerant of turbidity. Many farm dams and wetlands have turbidities of less than 135 NTU and the results of this experiment show that light is not the main factor limiting submerged plant establishment.

Since some water plants could establish under turbid conditions, a biological and chemical manipulation experiment was undertaken over 8 months in the Northern Tablelands of New South Wales, Australia. It aimed to establish water plants in farm dams and 'switch' them from a turbid state to a clear, water plant dominated state and test the hypothesis that clearing was not necessary for submerged plant establishment. Twelve turbid dams up to 3 m deep with no submerged or floating plants and without fish were chosen. Dams were fenced and domestic stock excluded or allowed only limited access.

Glasshouse trials were carried out on farm dam sediments both before and after the treatments, which showed that most farm dams had poor quality seed banks with few seedlings emerging before the treatments. Even though more species of water plants germinated in the glasshouse after the treatments, this increase also occurred in the control groups, which suggested a seasonal cause. Protection from grazing animals in the glasshouse also contributed to more species being observed there than in the field.

Laboratory tests were performed to determine the best chemical with which to flocculate suspended soil and algae. It was found that when lime flocculated clay and green algae in field and laboratory situations, it did not flocculate cyanobacteria (blue-green algae), as aluminium sulphate did. Aluminium sulphate was known to be toxic to aquatic animals and water plants so it was decided to use lime as the flocculating agent. Introduced seed bank collected from the temporary wetland (used in the previous pond trial) was used as the biological treatment as it contained turbidity tolerant charophytes and angiosperms. This seed bank was chosen because it did not have seeds of *Vallisneria* or *Typha*, species that under certain circumstances may dominate the vegetation of farm dams.

In early spring when green algae started to increase in abundance experimental treatments were applied to the farm dams. The treatments were either addition of lime (chemical treatment) or addition of seed bank (biological treatment), a combination of both lime and seed bank (chemical and biological treatments), and a control in which no treatments were applied (control). Three dams were used as controls. Water quality variables and submerged plant establishment was monitored for four months before and four months after treatment. The increase in the clarity of the water (as secchi disc depth) was measured. By introducing seed bank material to the dam sediments it was anticipated that submerged water plants would germinate and establish with increased light. Results showed that 4 months after treatment there was a significant difference in secchi disc depth in the lime and seed bank—lime treated dams but not in the control and seed bank treated dams. Chlorophyll_{-a} concentrations were reduced in all treatments but increased in the control group. The addition of seed bank did not increase submerged plant establishment in 6-8 weeks in the field as occurred in the artificial ponds. Grazing by ducks and aquatic animals

was detrimental to water plants and although water clarity improved in some dams there was no corresponding increase in submerged water plant germination. Lime treatment did not sustain water clarity in the long-term and high turbidity could indicate upstream erosion even where catchments are well-protected, not just wind and wave resuspension of sediments. Appropriate methods for protection from grazing birds and animals needs to be established as well as increased soil erosion control upstream, all which affected submerged plant establishment.

Where seed bank was used, colonization of the dam edges by emergent species was visible where slope was low but was not investigated. In contrast, submerged species were not successful in establishment. Long dry periods and the steep slopes of many dams led to large fluctuations in depth and did not support the establishment of any but the more hardy *Cyperus*, *Persicaria*, *Portulaca* and *Chenopodium* species and the naturalized couch and paspalum grasses.

Twenty months after treatment the secchi disc depth in the seed bank-lime and the seed bank treatment groups were higher than in the lime and control treatment groups. Unfortunately the averages were affected by stock disturbance to two dams of the previous two groups. In only one seed bank treated dam was there any large increase in submerged water plant germination. This dam had a 100% cover of *Nitella sonderi* to 0.54 m with a drop in turbidity from 57.8 NTU (av.) before treatment to 15.4 NTU; corresponding to an increase of secchi depth of greater than 1 metre. Soluble reactive and total phosphates were reduced in this dam by 50%/ 40% and 94%/ 77% four months and 20 months respectively after treatment. The establishment of submerged water plants in one dam out of six treated with seed bank may have occurred by chance alone, so it is uncertain if the addition of seed banks containing oospores of turbidity tolerant charophyte species was the mechanism which produced the 'switch'.

The long-term results of this project have shown that water plants can establish in turbid situations and at shallow depths so it is not necessary to clear the water to enable them to germinate. Farm dams can indicate the state of catchments and highly turbid dams indicate upstream erosion is occurring and control of this erosion must be a priority before water clarity can start to improve. Landcare groups can monitor the clarity of farm dams after rainfall events using secchi discs that would help them manage upstream soil erosion turbidity. Algal blooms may be natural and seasonal occurrences but when the blooms are excessive they can indicate a build up of nutrients in a water body. Lime can be used to manage these algal blooms in dams and postpone succession by blue-green algae, but this is a treatment for which long-term outcomes are unknown. There was an increase in water clarity in one of the seed bank treated dams in which charophytes established but this is in

no way conclusive as it happened outside the time frame of the project. Further research should be done on establishment of water plants, as blue-green algae remain a serious problem in Australian farm dams. Charophytes are small and low growing and provide habitat for algae consuming zooplankton and these three characteristics make them ideal for use in farm dams.

Table of Contents

FRONTISPIECE	1
DECLARATION	2
ACKNOWLEDGMENTS	3
ABSTRACT	Ę
TABLE OF CONTENTS	g
FIGURES	13
TABLES	16
PLATES	18
CHAPTER 1 LITERATURE REVIEW	19
1.1 The world-wide increase in water turbidity and the decline in submerged water plants	19
1.2 Definition of turbidity	20
1.3 Causes of turbidity in Australia	20
1.4 Turbidity and underwater light	23
1.5 Effects of turbidity on freshwater ecosystems	23
1.5.1 Physical effects of turbidity	23
1.5.2 Aquatic plant loss or change induced through increased turbidity	24
1.5.3 Aquatic plant losses through indirect turbidity effects	26
1.6 Effects of water plants on turbidity	26
1.7 Decline in submersed water plants in Australia	27
1.7.1 Historical	27
1.7.2 Recent	28
1.8 Increase in blue-green algae in Australian waters	29
1.8.1 Effects of physical changes in the water body on blue-green algae	29
1.8.2 Ecology of blue-green algae	31
1.8.3 Summary of blue-green algal survival skills	31
1.9 A local scale problem: New England Region of New South Wales	33
1.10 Water plant restoration from an Australian perspective	34
1.11 The alternative stable state theory	34
1.12 Aims and Thesis outline	37
PART 1	38
CHAPTER 2 EFFECTS OF FLOODING AND TURBIDITY ON WATER PLANT GERMINATION AND ESTABLISHMENT FROM WETLAND SEED BANKS	39
2.1 Introduction	39
2.2 Water plants	39
2.2.1 Water plants and flooding	39
2.2.2 Seed bank	40
2.2.3 Functional groups	40
2.3 Aims	41
2.4 Methods	43

1		٦
1	1	,

2.4.1 Experimental design	43		
2.4.2 Study sites and sources of seed bank	43		
2.4.3 Installing pots with seed bank in ponds	44		
2.4.4 Installing pots with seed bank in ponds	45		
2.5 Description of turbidity generation and experimental design	46		
2.5.1 Turbidity generation	46		
2.5.2 Setting up turbid ponds	47		
2.5.3 Testing the consistency of turbidity readings at depth	48		
2.6 Data collection:	48		
2.6.1 Light irradiance	48		
2.6.2 Physical parameters	52		
2.6.3 Water plant collection, identification and measurement	53		
2.7 Data analysis	54		
2.7.1 Measures of light and physical parameters	54		
2.7.2 Biological parameters – Germinations and Biomass	55		
2.7.3 Germination and biomass of charophytes and other species under different treatments and			
depths	55		
2.7.4 Lengths	56		
CHAPTER 3 RESULTS: EFFECTS OF FLOODING AND TURBIDITY ON WATER PLANT GERMINATION AND ESTABLISHMENT FROM POND TRIALS WITH WETLAND SEED BANKS	57		
3.1 Environment of ponds: light, chemical and physical parameters	57		
3.2 Water plants	61		
3.2.1 Functional species from two wetlands	61		
3.2.2 Species richness, germination and biomass	63		
3.2.3 Effect of depth on germination and biomass	68		
3.2.4 Effect of turbidity on the length of water plants	69		
3.2.5 Effect of turbidity on Myriophyllum variifolium	70		
3.2.6 Groupings of pots according to species determined by depth and turbidity	72		
CHAPTER 4 DISCUSSION: EFFECTS OF FLOODING AND TURBIDITY ON WATER PLANT GERMINATION AND ESTABLISHMENT FROM POND TRIALS WITH WETLAND SEED BANKS			
4.1 Introduction	77		
4.2 The underwater environment in ponds	77		
4.2.1 The suitability of using secchi depth as a measure of Z_{eu}	78		
4.2.2 Light irradiance in turbid ponds and the survival of water plants	80		
4.3 Effects of turbidity on species richness, biomass and germination of water plants	81		
4.3.1 Racecourse Lagoon seed bank	82		
4.3.2 Dumaresq Dam seed bank	83		
4.3.3 Use of seed banks in the rehabilitation of dams	85		
4.3.4 Effect of depth and turbidity on length	86		
4.4 Summary	87		
PART 2	91		
CHAPTER 5 FARM DAM REHABILITATION	92		
5.1 Introduction	92		
5.1.1 Reasons for low submerged plant establishment in farm dams	94		
5.1.2 Rehabilitation techniques or 'impacts'	96		

5.1.3 The use of lime in turbidity reduction	99
5.2 Aims	100
5.3 Methods	101
5.3.1 Study sites and experimental design	101
5.3.2 Treatments or impacts	102
5.3.3 Pre-treatment flocculation experiment	105
5.3.4 Discussion of treatment	106
5.4 Data collection	107
5.5 Glasshouse trials to detect the presence of water plant seeds in farm dam sediments	109
5.6 Data analysis	109
5.6.1 Sediment seed bank potential percentage cover and species richness.	109
5.6.2 Changes in measured water variables	110
CHAPTER 6 FARM DAM REHABILITATION: RESULTS	112
6.1 Introduction	112
6.2 Glasshouse farm dam sediment trials	112
6.3 Comparison of water variables (averages) of treatment groups before and after treatments	115
6.3.1 Secchi disc transparency	116
6.3.2 pH, Temperature, SRP and TP	120
6.3.3 Depth variation	121
6.4 BACI Analysis: Results of t-tests on water variables of treatment groups.	122
6.5 Biological changes	130
6.5.1 Plant germination in farm dams	130
6.5.2 Algal abundance as determined by chlorophyll- _a concentration	131
6.5.3 Zooplankton and Phytoplankton.	132
CHAPTER 7 FARM DAM REHABILITATION: DISCUSSION	134
7.1 Glasshouse germination of water plants from farm dam sediments:	134
7.2 BACI analysis: interpretation of the means, t-tests and significance of the changes in water vari of the treatment groups.	ables 136
7.3 Germination and establishment of water plants in farm dams	139
7.4 Zooplankton, Phytoplankton and relationship to chlorophyll- $_{a}$ concentration	141
7.5 Limitations of experiment	142
7.5.1 Sampling errors	142
7.5.2 Limitations to the interpretation of results	142
PART 3	144
CHAPTER 8 SYNTHESIS, MANAGEMENT RECOMMENDATIONS AND FUTURE RESEARCH	145
8.1 Synthesis	145
8.2 Management recommendations	147
8.3 Future research	149
REFERENCE LIST APPENDIX I	151 170
APPENDIX II	171
APPENDIX III	172
VI I PITAN 444	1,2

	12
APPENDIX IV	173
APPENDIX V	178
APPENDIX VI	179
APPENDIX VII	180
APPENDIX VII	181
APPENDIX VIII	184
APPENDIX IX	197

Figures

Downwelling irradiance, Ed (PAR) in μ mol m $^{-2}$ s $^{-1},$ measured in four waters of Figure 3-1 differing turbidities (clear and low NTU represented as light dashed and continuous lines and medium and high NTU represented as bold dashed and continuous lines) at three depths from the water surface. Top graph = 5 cm, Middle graph = 10 cm, Bottom graph = 20 cm. Light was measured around 59 noon when light intensity was most directly downward. Figure 3-2 Downwards irradiance, Ed (PAR) (µ mol m⁻² s⁻¹) measured at three depths, 5, 10 and 20 cm, for the four treatment groups (2 ponds per treatment). Turbidities 1.9 NTU = clear, 31 NTU = low, 70 NTU = medium, 140 NTU = high. Each point represents the mean of two ponds. Each pond was measured on 12 separate occasions twice weekly over a period of 6 weeks. An exponential trendline has been fitted to each series. Complete data is given in Appendix V 60 Figure 3-3 Average secchi disc transparency (Z_{SD}) in metres in paired ponds with varying turbidity indicating stability of visual readings at higher turbidities. Z_{SD} was maintained with clay concentrate. For Treatment 1 (control) Clear water, Z_{SD} > 1m (= depth of pond) so line not shown. (Assume 10's of metres (Kirk, 1994)). σ = Treatment 2, low turbidity; ϕ = Treatment 3, medium turbidity; \otimes = Treatment 4, high turbidity. For simplicity standard deviations are not shown on the graph but were ± 0.05 m, in the low, ± 0.02 m, in the medium and ± 0.01 m. in the high turbidity ponds. Increase in secchi depth over time in the low turbidity may be due to flocculation of suspended solids by bacteria, algae or 61 plants. Figure 3-4 Mean number of species which germinated from two seed banks from (a) Racecourse Lagoon (top) and (b) Dumaresq Dam (bottom) under three depth (5,10 and 20 cms) and four turbidity treatments (1) clear, 2) low, 3) medium and 4) high). Error bars represent standard deviations from the mean. 64 Figure 3-5 Germination (plants m²) and dry biomass (g m²) for Dumaresq Dam (a, b) and Racecourse Lagoon (c, d) seed banks at 5cm, 10cm and 20 cm depths for the four experimental turbidity treatments (1 = clear (control), 2 = low, 3 = medium,4 = high). Charophyte species are indicated by the height of the clear bars and all other submerged species are indicated by the total height of the gray bars. Error bars represent S.E.M. Note differences of scale between Dumaresq Dam 65 and Racecourse Lagoon. Figure 3-6 Changes in the length of different submerged plants at 20 cm depth with turbidity (1 = 1.9, 2 = 31, 3 = 70, 4 = 135 NTU). (a) Nitella subtilissima, (b) Nitella sonderi, (c) Chara fibrosa (d) Myriophyllum variifolium (e). Chara muelleri. The seed bank was collected from Racecourse Lagoon. Average length in cm and the SEM are shown on top of columns. 71 Figure 3-7 Dendrogram showing the grouping of pots containing Racecourse Lagoon seed bank from different depths and turbidity treatments after cluster analysis using PATN (Belbin, 1991). Pots were clustered according to the species and numbers of germinations from the seed bank. Left hand number 1 to 8 represents treatment pond (see below); R =Racecourse Lagoon; A, B or C represents 5, 10 or 20 cm depth respectively; right hand number 1 or 2 represents replicate in pond. Ponds 1 & 2 = Clear water treatment, ponds 3 & 4 = low turbidity treatment, Ponds 5 & 6 = medium turbidity treatment, Ponds 7 & 8 = high turbidity treatment. 75

rigure 3-8	from different depths and turbidity treatments after cluster analysis using PATN (Belbin, 1991). Pots were clustered according to the species that germinated and established from the seed bank. Left hand number represents treatment pond (see below); D = Dumaresq Dam; A, B or C represents depth 5, 10 or 20 cm respectively; right hand number 1 or 2 represents replicate in pond. Treatment ponds 1 & 2 = Clear water treatment, Ponds 3 & 4 = low turbidity treatment, Ponds 5 & 6 = medium turbidity treatment, Ponds 7 & 8 = high turbidity treatment.	76
Figure 5-1	Diagram showing the five groups of dams identified in Casanova et al. (1997) as they relate to the 4 phases identified by Moss et al. (1996). Phase I - aquatic plant dominant state: with abundant water plants, high water clarity, low nutrient concentrations, low phytoplankton nos. Phase II - few water plants, low to medium nutrient concentrations, low phytoplankton nos. Phase III- no water plants, medium to high nutrient concentrations, moderate phytoplankton nos. Phase IV- phytoplankton dominant state: with high phytoplankton nos. (usually blue-green algae), an absence of water plants, high nutrient concentrations, and high turbidity. Phase V- amenity dam with introduced, floating water plants. Phases I & IV are described as being in 'alternative stable states', Phases II and III are described as 'transition phases'. Bold arrows represent 'forward switches'; dotted arrows represent 'reverse switches'.	93
Figure 5-2	Boundaries of the area for the farm dam rehabilitation experiment in the Macleay River catchment of the New England Tablelands and location of Landcare groups. Dams are shown as black dots. Scale 2 cm = 20 km. Northwestern boundary of marked area is approximately the western edge of the Great Dividing range.	104
Figure 6-1	Glasshouse trials on farm dam sediment seed bank germination in relation to experimental treatment: (a) Average number of submerged water plant species that germinated from each dam sediment and (b) average percentage cover of trays. Results for (a) and (b) are for pooled depths 5 and 20 cm after 3 months. Error bars represent one standard deviation.	115
Figure 6-2	Results from a Before-After-Control-Impact-pairs (BACIP) experiment to see if the secchi disc transparency depth changed over time after treatment (impact) with either seed bank, lime or seed bank-lime. For each point $n=3$. Control group was not treated. Treatment (shown by arrow) was done directly after 4^{th} sampling in October 1997. Standard deviations from group means are not shown on this graph for simplicity; for standard deviations see Figure 6.3.	119
Figure 6-3	Results from a BACIP experiment to see if the secchi disc transparency depth (secchi depth) changed over time after treatment (impact) with either seed bank, lime or seed bank-lime. Treatment was after October 1997 (4th sampling). Note difference from Figure 6.2 20 months after treatment (9th sampling) (June 1999) and standard deviations in seed bank treatments due to introduction of stock to one of the three replicated dams. Arrow shows time of treatment. Error bars represent one standard deviation.	119
Figure 6-4	Secchi depth in individual dams before and after treatment and 20 months after treatment. C = control, SB = seed bank, L = lime, SBL = seed bank-lime treatment. Numbers represent dam number. 'X' over columns represents dams that were impacted by stock introduction. Error bars represent one standard deviation for the average secchi depth in each dam. No standard deviation is given for the column '20 months after' or the 9 th sampling, as it was a 'one-off' sampling.	120

igure 6-5	Results from a BACIP experiment to see whether (a) soluble reactive phosphate (SRP) and (b) total phosphate (TP) both in mg L ⁻¹ of four treatment groups: control, seed bank, lime and seed-bank-lime changed after treatment. Treatment time (shown by arrow) was after the 4 th sampling in mid-October 1997.	130
Figure 6-6	Results from a BACIP experiment to see if the chlorophyll- _a concentration in four treatment groups: control, seed bank, lime and seed bank-lime changed after treatment. Treatment time (shown by arrow) was after the 4 th sampling in mid-October 1997. Two samplings for chlorophyll- _a (July and August) were not	122
	taken. Error bars represent one standard deviation from the mean.	132

Tables

Table 1 1	Causes and effects of increased abiotic and biotic turbidity in Australian waters, rivers and wetlands. NB: Some of these references may quote several causes and effects	21
Table 2 1	Species of submerged and attached floating water plants found in two wetlands: Racecourse Lagoon and Dumaresq Dam (Casanova 1993). 45	
Table 2 2	Rule of thumb for generating turbidity (NTU) using clay stock (approximate)	47
Table 2 3	List of physical, chemical and biological measurements used in Chapter 2-4 (pond experiment) and Chapter 5-7 (farm dam experiment and the sediment germination experiment in the glasshouse.	50
Table 2 4	Measurements, symbols and units commonly used in light studies.	52
Table 3 1	Initial readings of SRP and NO ₃ -N and initial and final pH in all turbidity treatments. (Average of two samples – no variation. 58	
Table 3 2	Results of average turbidity, secchi depth (Z_{SD}) ; total incident radiation flux, [Ed (z) +Eu (z)], euphotic depth (Z_{eu}) (calculated) and temperature (^{0}C) in four turbidity treatments in ponds	61
Table 3 3	Species of water plant common to two seed banks which germinated and established in a range of turbidities from clear to high (0-140 NTU) at three depths (5, 10 and 20 cm)	62
Table 3 4	Average number of <i>Vallisneria gigantea</i> plants m ² (converted from plants per pot by multiplying by a factor of 44) that established from Dumaresq Dam seed bank under three depth and four turbidity treatments (clear, low, medium high). Numbers include vegetatively produced clones of parent plant	69
Table 3 5	Reduction in the average total dry biomass grams per pot (diam 17 cm) of <i>M. variifolium</i> grown from two seed banks, Racecourse Lagoon and Dumaresq Dam, when placed at three depths; 5, 10 and 20 cm underwater (using data pooled from four turbidity treatments). (Different numbers of plants were weighed from each wetland but there was a similar reduction in biomass with depth.)	72
Table 5 1	Turbid farm dams and their responses to two chemical flocculation treatments. Treatments of dams were chosen at random and then 4 X 500ml water was tested for its response to both aluminium sulphate (alum) and calcium hydroxide (lime) stock solutions. (Stock 1 mgml ⁻¹). Dam 2 flocculated in the field with lime. **Dam 5 had a blue green algal bloom at the time, which only flocculated with alum.	105
Table 5 2	Treatment impact on dams and quantities of lime and seed bank used. Each treatment (Impact) was randomly allocated to each of the four dams in the same neighbourhood (group). Seed bank amounts are calculated from buckets of seed bank spread by hand. 1-bucket seed bank covers 5 m^2 to 1-cm depth. *Note- Dam 5 with blue-green algae; algae was not flocculated by lime although treatment was still given. Impact (treatment) $C = \text{control}$, $SB = \text{seed bank}$, $L = \text{lime}$, $SB/L = \text{seed bank}$ and lime.	105
Table 6 1	Species which germinated in the submerged trays (pooled 5 and 20 cm depth) of farm dam sediments before (spring) and after (autumn) treatment. Treatments: $Ctrl = control$, $Sb = seed bank$, $L = lime$, $Sb-L = seed bank-lime$. $B = before treatment$, $A = after treatment$.	113

Table 6 2	Averages of water quality variables for four groups of dams in which parameters were measured for 4 months before and 4 months after impact (various treatments) and their difference. $n=12$ for both before and after, except for chlorophyll- $_a$ where $n=6$ Before and $n=12$ After. Impacts (treatments) were: Control (no treatment), seed bank, lime and seedbank-lime. A postive sign indicates an increase in the parameter, a negative sign indicates a decrease. Secchi depth (Secchi) in m., turbidity in NTU, conductivity (Conduct.) in μ S cm $^{-1}$, chlorophyll- $_a$ (Chloroph.) in μ g L $^{-1}$, SRP and TP in mg L $^{-1}$, temperature (Temp) as degrees centigrade, $^{\circ}$ C.	118
Table 6 3	Comparisons of Secchi Disc Depth among four treatment groups before and after treatment with seedbank (2), lime (3) and seedbank and lime (4). A fourth group was a control (1). The significance of differences was tested using a two sample t-tests. Degrees of Freedom (DF), T and level of significance (P) are shown for each comparison.	122
Table 6 4	Comparisons of turbidity among four treatment groups before and after treatment with seed bank (2), lime (3) and seedbank and lime (4). A fourth group was a control (1). The significance of differences was tested using a two sample t-tests. Degrees of Freedom (DF), T and level of significance (P) are shown for each comparison.	123
Table 6 5	Comparisons of conductivity among four treatment groups before and after treatment with seedbank (2), lime (3) and seedbank and lime (4). A fourth group was a control (1). The significance of differences was tested using a two sample t-tests. Degrees of Freedom (DF), T and level of significance (P) are shown for each comparison.	124
Table 6 6	Comparisons of pH among four treatment groups before and after treatment with seedbank (2), lime (3) and seedbank and lime (4). A fourth group was a control (1). The significance of differences was tested using a two sample t-tests. Degrees of Freedom (DF), T and level of significance (P) are shown for each comparison.	125
Table 6 7	Comparisons of temperature among four treatment groups before and after treatment with seedbank (2), lime (3) and seedbank and lime (4). A fourth group was a control (1). The significance of differences was tested using a two sample t-tests. Degrees of Freedom (DF), T and level of significance (P) are shown for each comparison.	126
Table 6 8	Comparisons of soluble reactive phosphate (SRP) among four treatment groups before and after treatment with seedbank (2), lime (3) and seedbank and lime (4). A fourth group was a control (1). The significance of differences was tested using a two sample t-tests. Degrees of Freedom (DF), T and level of significance (P) are shown for each comparison.	127
Table 6 9	Comparisons of total phosphate(TP) among four treatment groups before and after treatment with seedbank (2), lime (3) and seedbank and lime (4). A fourth group was a control (1). The significance of differences was tested using a two sample t-tests. Degrees of Freedom (DF), T and level of significance (P) are shown for each comparison.	128
Table 6 10	Comparisons of chlorophyll- _a among four treatment groups before and after treatment with seedbank (2), lime (3) and seedbank and lime (4). A fourth group was a control (1). The significance of differences was tested using a two sample t-tests. Degrees of Freedom (DF), T and level of significance (P) are shown for each comparison.	129
Table 6 11	Average number of zooplankton and phytoplankton taxa and their standard deviation in groups of farm dams before and after treatment and the change in species richness. Treatments: Control, seed bank, lime and seed bank-lime.	133

Plates

Plate 1	(a) Artificial ponds of turbid water used for wetland seed bank germination trials.	203
	(b) Turbidity tolerant charophyte species (a) Chara muelleri (b) Nitella subtilissima (c) Chara fibrosa and (d) Nitella sonderi.	
	(c) Charophytes grown in a pot at 20 cm in a highly turbid pond of 135 NTU.	
Plate 2	(a) Turbid farm dam on granite soil with no submerged water plants – one of the many thousands of farm water storages in Australia which have replaced natural wetlands.	204
	(b) Spreading lime on a farm dam from a mixing tank	
	(c) Collection of seed bank from wetland	
	(d) Drying bags of seed bank	
	(e) Sieved and mixed seed bank ready for spreading	
Plate 3	Trays of sediment from farm dams placed in clear water in glasshouse to investigate submerged water plant germination.	205
Plate 4	Establishment of emergent species from introduced seed bank on different slopes.	206
	(a) Slope low and suitable for emergent water plant establishment and reproduction.	
	(b) Slope too high with emergent water plants left high and dry after a decrease in water level. Sexual and vegetative reproduction cannot be completed.	
	(c) Results of wood duck (<i>Chenonetta jubata</i>) grazing on shallow water and amphibious vegetation at edge of farm dam after seed bank addition.	
Plate 5	(a) University of New England and Landcare field day on 'Water quality in farm dams' at 'Terrible Vale', Kentucky and 'Wilmenshoe', Uralla	207
	(b) Demonstrating secchi disc transparency depth for submerged plant establishment.	
Plate 6	Poster: 'The Role of Turbidity in Water Plant Establishment' presented at ASL 1996	208