CHAPTER 7

GENERAL DISCUSSION

The studies described in the experimental Chapters of this thesis, focus primarily upon the improvement, by genetic means, of the reproduction rate of the Merino breed in Australia. This industry is characterised by low lambing rates when compared to sheep breeding industries in many other countries. Bindon and Piper (1976) have identified the major limiting factors which restrict improvement in reproduction rate in Merino flocks in Australia, and among the most important of these are the inherent low prolificacy of the Merino breed and the high emphasis, in selection programmes, which is placed on the maintenance and improvement of production of wool free from contamination by pigmented fibres. The great majority of the prolific breeds found throughout the world are characterised by either coarse or pigmented fibres, and this factor has largely ruled out improvement of the prolificacy of Merino flocks by crossbreeding with exotic breeds.

The other major factor which hinders the improvement of reproduction rate in Merino flocks is the environment. The majority of the breeding flocks in Australia are found in the relatively low rainfall areas where control of nutrition of the breeding flock is often achieved through manipulation of stocking rate rather than by supplementary feeding. Not only does the climate limit the expression of "potential" ovulation rate, and hence prolificacy, but also limits the opportunities for exerting adequate selection pressure on the female.

With these factors in mind, the concept advanced by Land (1973), that selection for prolificacy in sheep populations may be enhanced by utilising male "marker" traits which are genetically correlated with female reproductive performance, has particular relevance to genetic improvement of reproduction rate in the Australian Merino industry.

The studies described in Chapters 3, 4 and 5 have aimed at quantifying, in a large random breeding Merino flock, the genetic parameters required to accurately assess whether the use of male reproductive traits as indirect selection criteria for female reproductive performance would be more efficient than the methods currently in use.

Given our lack of knowledge of the factors influencing variation in male reproductive traits in Merino flocks maintained under pastoral conditions, it was considered important to first identify and quantify these sources for the male traits under study. Testicular diameter was chosen as the primary trait to be investigated because of its clear genetic association with female reproductive traits in laboratory animals and also because of its ease of measurement.

The results of the study of testicular diameter in the Trangie D flock, reinforce the conclusions from other studies that the trait is moderately to highly heritable. In addition, the genetic correlations estimated from the repeated monthly measurement data, suggest that during the period from 5 to 12 months of age, the expression of testicular diameter is controlled largely by the same set of genes. Allowing for the fact that the heritability of the trait apppeared to be lower at 5 months of age, which was thought to be due to masking by maternal influences, the results suggest that ram selection on

testicular size could be successfully practiced from 6-7 months of age. However, it was not until the rams were aged 9-10 months that the penalty of being born as a twin or triplet became non-significant. Therefore ram selection indices for flocks maintained under similar nutritional conditions to those pertaining to this study and which include testicular diameter as a criterion, should ideally utilise a measure taken between 10 and 12 months of age.

The phenotypic and genetic relationships between liveweight and testicular diameter were positive at all ages in the study. However, liveweight at weaning age was not closely correlated, phenotypically and genetically, with testicular size at later ages. Evidence was also found from the examination of testicular diameter at specific liveweights, that at a liveweight typical of an immediate pre-weaning stage of development (17kg), testicular diameter was lowly and sometimes negatively correlated with testicular diameter at higher liveweights. These findings suggest, again, that selection decisions using testicular diameter as a criterion may be better delayed until at least 10 months of age.

The study of serving capacity in the 20-month old rams of the Trangie D flock was highlighted by the absence of additive genetic variation in the measures of sexual performance studied. The relative instability of sexual performance, at both ends of the scale, suggests that the provision of 2 introductory tests of 20 minutes duration was insufficient to allow the establishment of stable behavioural patterns. Further studies of the effect of heterosexual experience prior to testing of young rams for serving capacity, are required to establish the effect of such treatment on both the incidence of sexual

inhibition and on adult levels of serving performance. There also remains the necessity for further investigations of the relationship between pen- and yard-test performance and paddock-mating performance of young rams. Such studies, however, may have to await the further development of remote electronic sensing technology for more labourefficient monitoring of paddock mating activity.

Genetic relationships between male and female reproductive traits were the subject of the study reported in Chapter 5. Estimates of genetic correlations between testicular diameter and ovulation rate at various ages were derived using dam-son and paternal half-sib covariance methods. In the former case, the method required the use of genetic variances which were not derived from exactly the same population as were the covariance estimates. However, by regarding the correlation estimate, at each age, as a small sample estimate, and pooling across ages, where appropriate, more precise estimates were obtained. Likewise, with the paternal half-sib REML method, starting values of genetic variances and covariances for this iterative procedure, were derived from closely related but not identical populations. It was possible, however, by a pooling procedure to arrive at one overall estimate of the half-sib genetic correlation between testicular diameter at 5-12 months of age and ovulation rate at 18-54 months of age.

The overall pooled values of 0.35 ± 0.08 and 0.16 ± 0.11 estimated by the dam-son and paternal half-sib covariance methods, respectively, differ substantially in their practical impact when used in simple selection response equations. When considering the implications of these 2 estimates, it is relevant to note that in comparison to the

half-sib estimate, that calculated from the dam-son covariances includes a larger contribution due to possible epistatic effects. In selection schemes, where the sire-daughter covariation will be the medium through which selection will act, epistasis of similar magnitude could be expected.

From a practical viewpoint, the results of the studies described in Chapters 3 and 5 provide only the starting points for a detailed evaluation of the use of testicular size as a criterion in ram selection indices. The genetic parameters estimated in these studies will, however, allow a far more rational evaluation of the use of testicular size as a selection criterion for Merino flocks maintained under pastoral conditions than has been the case to date. Further studies of the economic benefits of increasing testicular size, from the viewpoint of "current flock" fertility are required.

Chapter 6 described investigations of the expression of the Booroola Merino \underline{F} gene in the male phenotype. Whereas the other Experimental Chapters in this Thesis have described studies of traits under the control of many genes of small effect, in Chapter 6 we were concerned with the expression of a gene which has been shown to have a major effect on the female. In retrospect, the expectation that the expression of the \underline{F} gene should be evidenced in both sexes as a deregulation of gamete release by the gonad, ignores fundamental differences in the endocrine characteristics of the two sexes. Given the differences between the sexes in the plasma concentrations of both gonadotrophins and steroids, it could be expected that there would exist substantial differences between the sexes in the threshold levels of hormones which are required to trigger germ and somatic cell differentiation and growth. Such differences could easily mask the effect of a gene in one sex, no matter how dramatic its effect in the other. In the extreme case, the differential effect of a gene in the two sexes could produce quite dissimilar endocrine profiles in peripheral plasma of male and female <u>F</u> gene carriers, when comparisons with non-carriers are made. Therefore the explanation for the differences between <u>F</u> gene carriers and non-carriers found in the study of LH profiles at 10 weeks of age, should not be argued from the basic knowledge we have from similar studies with females. Rather, this finding demands more detailed studies of males of equivalent genotypes around this critical age (and liveweight), with a view to fully characterising gonadotrophin and steroid profiles. Such investigations would also serve as a replicate of the study described in Chapter 6, thereby increasing the power of the comparisons between the genotypes.

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Yeates, N. T. M. 1949). The breeding season of the sheep with particular reference to its modification by artificial means using light. J. Agric. Sci. Camb. <u>39</u>: 1-43. APPENDIX 1.

Genetic variances $(\delta m^2 \text{ and } \delta f^2 \text{ for male and female traits,}$ respectively) of testicular diameter (TDM) at 5, 8, and 12 months of age and ovulation rate (OR) at 18, 30, 42, and 54 months of age and covariances between them (Cov_g), derived from REML analyses of data from the Trangie D flock half-sibs born in 1979-1981.

Trait TDM	pair OR	Year	δm^2	Covg	§f ²
5	18	1979	0.034282	0.005776	0.004476
		1 9 80	0.022147	0.034638	0.010479
		1981	0.071097	0.010848	0.008193
8	18	1979	0.202403	0.002570	0.003454
		1980	0.486429	-0.004956	0.011248
		1981	0.632233	0.000797	0.002562
12	18	1979	0.151749	0.010330	0.004746
		1 9 80	0.148747	0.011856	0.012692
		1981	0.199172	0.002488	0.008101
5	30	1979	0.042668	0.010198	0.023098
		1 9 80	0.198711	0.001211	0.007498
		1981	0.076242	0.032393	0.028281
8	30	1979	0.211558	0.015711	0.022173
		1980	0.486814	0.001615	0.004595
		1981	0.626195	0.013825	0.021845
12	30	1979	0.153156	0.010740	0.021900
		1 9 80	0.148130	-0.000064	0.004577
		1981	0.112575	0.027664	0.022040

APPENDIX 2

ANOVA mean squares for morphological and hormonal traits from rams in

Experiments 1 and 2

Source	df	Pituitary weight(g)	Testicular weight(g)	Epididymal weight (g)	Live weight (kg)	Testicular diameter(cm)	Plasma FSH (ng/ml)	Plasma LH (ng/ml)
Experiment	1	0.055633**	23570.179***	75.9376***	8930.215***	2.937716**	0.015451	4.090050
Strain	1	0.005971	840.364	23.1358	29.179	0.099835	0.086723	0.003286
Dam genotype w/n Booroola	1	0.001712	266.354	47.5541	0.493	0.544665	0.067526	0.157097
Sire	11	0.006974	1972.466	26.8332	101.206	0.343545	0.022369	0.061308
Birth type	3	0.005116	102.140	7.5904	7.701	0.144739	0.059997	0.046409
Remainder	26	0.004403	1098.844	26.1629	91.311	0.307811	0.071111	0.124077

Trait

APPENDIX 3

ANOVA mean squares for TDM, and mean plasma FSH and LH at 7 ages from rams in Experiment 2

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	Testicular diameter (cm) at age:							
Source	df	3 weeks	5 weeks	10 weeks	14 weeks	18 weeks	26 weeks	33 weeks
Strain	1	0.002447	0.031817	0.228285	0.094314	0.049025	0.159703	0.479318
Sire	4	0.040051	0.065461	0.103356	0.204612	0.116077	0.176440	0.379898
Birth type	2	0.089769*	0.164915*	0.112930	0.651576*	0.095210	0.186624	0.183050
Booroola dam genotype	1	0.022055	0.086243	0.067245	0.536150	0.051444	0.401986	0.575304
Remainder	14	0.019023	0.028913	0.125421	0.172312	0.158467	0.131738	0.291528
	Mean plasma FSH (ng/ml) at age:							
Source	df	3 weeks	5 weeks	10 weeks	14 weeks	18 weeks	26 weeks	33 weeks
Strain	1	0.070171	0.008369	0.002104	0.043969	0.010942	0.040102	0.007487
Sire	4	0.349775	0.409904	0.259969	0.130922*	0.019431	0.350582	0.026229
Birth type	2	0.285315	0.447656	0.191938	0.131646	0.011033	0.047001	0.038872
Booroola dam genotype	1	0.058923	0.486938	0.407523	0.044503	0.002886	0.275457	0.018037
Remainder	14	0.203644	0.207358	0.166852	0.041891	0.009004	0.207335	0.100989
Mean plasma LH (ng/ml) at age:								
Source	df	3 weeks	5 weeks	10 weeks	14 weeks	18 weeks	26 weeks	33 weeks
Strain	1	0.025259	0.250646	3.900597	0.058540	0.033355	1.304325	0.101010
Sire	4	0.072307	0.205924	3.519880	0.719883	0.268059	0.670164	0.075880
Birth type	2	0.120910	2.018632	2.889191	0.003562	0.040350	2.307850	0.024174
Booroola dam genotype	1	0.002983	1.649170	12.109722*	0.189980	0.724204	0.008837	0.008920
Remainder	14	0.267907	0.579389	1.574955	0.515812	0.237023	1.102139	0.193932