Effect of root diseases and drought on water use efficiency of wheat

By

Abdallah Aldahadha

B.Sc.: Plant Production (Jordan University of Science and Technology, Irbid, Jordan)

M.Sc.: Horticulture (Jordan University, Amman, Jordan)

A thesis submitted for the degree of

Doctor of Philosophy

At The University of New England

Botany

School of Environmental and Rural Sciences

Armidale, NSW, 2351

Australia

Acknowledgements

I would like to express my gratitude to my principal supervisor, Dr. Nigel Warwick and cosupervisor, Dr. David Backhouse, for their excellent guidance and support during my study at the University of New England. I would like to thank them for their patience when teaching me different techniques in the areas of plant ecophysiology and pathology and statistical analysis. I am most grateful for their invaluable suggestions during the writing up of this thesis and their support to attend several conferences.

I would like thank and acknowledge Mr. Richard Willis and Mr. Michael Faint for their help in the greenhouse, collecting soil and other technical support. I am grateful to Dr. Dale Kenny for his help in taking physiological measurements in the greenhouse. I also thank all academic staff in Botany for their kindness and friendship.

I wish to thank all my friends in the University of New England. Finally, the greatest thanks to my family for all kind of supports and assistance and for providing me with the mental strength throughout my studies.

Dedication

This thesis is dedicated to my lovely parents, sisters and brothers. I would like to thank all of them, particularly my mother, who provided me with support, love and encouragment to enable me to achieve my goals. I also thank all of them for the financial support which enabled me to complete my study.

Abstract

In arid and semi-arid areas, wheat production is limited by drought mainly at the post-anthesis stage. Root diseases also affect the productivity of wheat. This work reports an examination of the response of wheat plants to the interaction of drought and root diseases. The major hypothesis was that root diseases reduce water use efficiency (WUE) under drought conditions.

The preliminary experiment compared two cultivars, two root diseases and drought at two plant growth stages. There were no differences between the two root diseases, *Pythium* and *Rhizoctonia*, however, there was reduced transpiration in diseased plants of cv. Janz following tillering drought, and of cv. Mulgara following anthesis drought. WUE was not affected by diseases.

In experiments with deeper pots and post-anthesis drought treatment, *Pythium* reduced transpiration prior to anthesis. When plants were regularly rewatered to field capacity before the post-anthesis drought, *Pythium* had no effect on grain yield and WUE grain. When both infected and control plants were given the same total volume of water at high inoculum densities, infected plants had higher WUE grain than controls. Water uptake by infected roots was reduced during early plant growth due to root damage, but was higher than controls during post-anthesis drought due to higher remaining soil water availability, when compared with non-infected plants. However, diseased plants were not able to access all of the additional water and grain yields were not different from controls.

Hydroponic experiments compared the effects of *Pythium* and root pruning on plant water relations. *Pythium*, polyethyleneglycol-induced drought and root pruning reduced

transpiration to a similar extent, but only *Pythium* reduced shoot dry weight. *Pythium* had no effect on total or instantaneous WUE. This experiment suggested that the effect of *Pythium* was more than just in reducing the effective size of the root system. Root pruning at a late stage of vegetative growth in pots reduced WUE, demonstrating the importance of the stage at which root damage occurs on how it affects total WUE.

Overall, root diseases did not reduce WUE. Reduction in growth was due to reduced transpiration. This did, however, lead to more water being available in soil during a post-anthesis drought.

Declaration

I certify that the substance of this thesis has not already been submitted for any other degree or qualification.

I certify that any help received in preparing this thesis, and all sources used, have been acknowledged in this thesis.



Abdallah Aldahadha

Table of Contents

Acknowledgementsi
Dedicationii
Abstractiii
Declarationv
Table of Contentsvi
List of Tablesx
List of Figuresxi
Chapter 1: General introduction
1.1 Background
1.2 Research objectives
Chapter 2: Literature review
2.1 Introduction
2.2 Wheat
2.3 Drought effects on plant physiology
2.3.1 Yield and biomass production
2.3.2 Effect of drought on yield components
2.3.3 Yield and water use (evapotranspiration)
2.3.4 Water use efficiency 14
2.3.5 Plant water relations 19
2.3.6 Water uptake by roots
2.3.7 Other physiological effects
2.4 Root diseases effects
2.4.1 Pythium disease of wheat
2.4.2 Rhizoctonia disease of wheat
2.4.3 Root damage by fungal diseases
2.4.4 Yield losses from root diseases in wheat
2.4.5 Disease and water relations
2.4.6 Effect of diseases on physiological parameters in wheat
2.5 Drought X pathogen interaction in other plants
2.6 Conclusion

Chapter 3: Interactive effects of drought and fungal roof wheat	•
3.1 Introduction	
3.2 Materials and methods	39
3.2.1 Inoculum preparation	39
3.2.2 Soil preparation	39
3.2.3 Inoculation of wheat plants	40
3.2.4 Growth conditions and treatments	40
3.2.5 Measurements	41
3.3 Results	44
3.3.1 Variation of growth stages	44
3.3.2 Water Use (Transpiration)	46
3.3.3 Plant water relations	54
3.3.4 Yield components	58
3.3.5 Water Use efficiency (WUE)	60
3.3.6 Lesion percentage	
3.3.7 Qualitative effects of disease on root systems	66
3.4 Discussion	
Chapter 4: Effect of <i>Pythium</i> on water use efficiency an under drought	
4.1 Introduction	74
4.2 Material and Methods	76
Experiment 1: Effect of different inoculum density of <i>Pyth</i> yield components of wheat	•
4.2.1 Preparation of inoculum	76
4.2.2 Soil preparation and soil inoculation with Pythic	ım76
4.2.3 Water regime	77
Experiment 2: Effect of Pythium on water relations and ot	her physiological parameters 78
4.2.4 Soil preparation and treatments	
4.2.5 Measurements	79
4.2 Deculto	90

Experiment 1: Effect of different inoculum density of Pythium on yield and water use efficiency	80
4.3.1 Cumulative transpiration	80
4.3.2 Transpiration per week	81
4.3.3 Yield components	82
4.3.4 Water use efficiency	83
Experiment 2: Effect of <i>Pythium</i> on water relations and other photosynthetic parameter	s 84
4.3.5 Cumulative transpiration	84
4.3.6 Transpiration per week	86
4.3.7 Yield components	86
4.3.8 Water use efficiency (WUE)	87
4.3.9 Plant water relations	88
4.3.10 Physiological measurements	91
4.4 Discussion	100
Chapter 5: Effects of <i>Pythium</i> and root pruning on water use efficiency of hydroponically grown wheat under PEG-induced drought	108
5.1 Introduction	108
5.2 Materials and methods	110
5.2.1 Plant material and growth conditions	110
5.2.2 Root inoculation by <i>Pythium</i>	112
5.2.3 Root pruning and Polyethylenglycol (PEG)	114
5.2.4 Measurements	115
5.3 Results	117
5.3.1 Transpiration per day	117
5.3.2 Transpiration per week	119
5.3.3 Plant Growth	122
5.3.4 Water use efficiency	124
5.3.5 Plant water relations	126
5.3.6 Physiological measurments	128
5.4 Discussion	131
Chapter 6: Effect of Root Pruning on Water Use Efficiency of Wheat	137

6.1 Introduction	137
6.2 Materials and Methods	139
6.3 Results and Discussion	141
6.3.1 Transpiration	141
6.3.2 Grain yield and biomass components	144
6.3.3 Water Use Efficiency	147
Chapter 7: General Discussion and Future Directions	149
References	162
Appendix	185

List of Tables

Table 3.1 Number of tillers per pot (TN/pot), number of heads per pot (HN/pot), grain weight
(GW), dry matter weight (DMW), number of grains (GN), 1000-grain weight (1000-
GW) and harvest index (HI) for well watered (WW), tillering drought (D1) and anthesis
drought (D2) of two wheat cultivars under combined disease conditions
Table 4.1 Number of tillers per pot (TN/pot), number of heads per pot (HN/pot), grain weight
(GW), dry matter weight (DMW), and harvest index (HI) of wheat cv. Janz wheat
following inoculation with different levels of inoculum of <i>Pythium</i> (g/pot) (ID). Values
are means of 6 replicates
Table 4.2 Cumulative transpiration of wheat cv. Janz for controls and <i>Pythium</i> at three growth
stages
Table 4.3 The number of heads per pot (HN/pot), grain weight (GW), dry matter weight
(DMW), and harvest index (HI) for wheat cv. Janz after inoculation with 10 g of
<i>Pythium</i> per pot
Table 5.1 Dates of activities for hydroponic experiments 1 and 2
Table 5.2 Effect of root pruning and Pythium in the presence (+PEG) and absence (-PEG) of
polyethylene glycol on transpiration of wheat cv. Janz during 7 days in which PEG and
root pruning treatments were imposed. Each value represents the mean of eight
plantsper treatment
Table 5.3 Effect of root pruning and <i>Pythium</i> in presence (+) and absence (-) of PEG on root
and shoot dry weights, and root: shoot ratio of cv. Janz in first experiment. Each value
represents the mean ±SE of eight plants per treatment
Table 5.4 Effect of root pruning and <i>Pythium</i> in presence (+) and absence (-) of PEG on root
and shoot dry weights, and root/ shoot ratio of cv. Janz in second experiment 124
Table 6.1 Number of heads per pot (HN/pot), grain weight (GW), number of grains (GN),
shoot dry weight (SDW), root dry weight (RDW), root/shoot ratio (R:S ratio), harvest
index (HI) and 1000-grain weight (1000-GWt) for controls, infected and root pruned
plants of cv. Janz
•

List of Figures

Figure 2.1 The several means of water use efficiency. Source, Tambussi et al. (2007) 15
Figure 3.1 Effect of drought at tillering (D1) and disease on growth stage one week after
drought treamtent for Triticum aestivum cvs. Mulgara (M) and Janz (J). WW: well
watered, C: controls, P: Pythium and R: Rhizoctonia
Figure 3.2 Effect of anthesis drought (D2) and diseases on growth stages one week after
drought treatment for <i>Triticum aestivum</i> cvs. Mulgara and Janz . WW: well watered, C:
controls, P: Pythium and R: Rhizoctonia
Figure 3.3 Cumulative transpiration (mm) from 9 April (tillering) to 23 July (harvest) for cvs.
Mulgara (left) and Janz (right) under diseases treatments: Pythium (P), Rhizoctonia (R)
and control (C) and drought treatments at tillering (D1) and anthesis (D2) Upper: well
water, Middle: D1 and Lower:D2. 48
Figure 3.4 Transpiration per week (mm.week ⁻¹) from 9 April (tillering) to 23 July (harvest)
for cvs. Mulgara (left) and Janz (right) under disease treatments (Pythium (P),
Rhizoctonia (R) and control (C)) and drought treatments at tillering (D1) and anthesis
(D2). Upper: well watered, Middle: D1 and Lower:D2
Figure 3.5A Cumulative transpiration (mm) of well watered (WW) and droughted at tillering
(D1) of plants of cv. Mulgara under different disease conditions (Pythium P;
Rhizoctonia R; control c) during 7 days of water stress
Figure 3.5B Cumulative transpiration (mm) of well watered (WW) and droughted at tillering
(D1) of plants of cv. Janz under different disease conditions (Pythium P; Rhizoctonia R;
control c) during 7 days of water stress
Figure 3.5C Cumulative transpiration (mm) of well watered (WW) and droughted at anthesis
(D2) of plants of cv. Mulgara under different disease conditions (Pythium P;
Rhizoctonia R; control c) during 7 days of water stress
Figure 3.5D Cumulative transpiration (mm) of well watered (WW) and droughted at anthesis
(D2) of plants of cv. Janz under different disease conditions (Pythium P; Rhizoctonia R;
control c) during 7 days of water stress
Figure 3.6A Water potential (left), osmotic potential (mid) and pressure potential (right) in cv.
Mulgara (M) infected with Pythium (P), Rhizoctonia (R) and control (C). The
measurements were taken at the beginning and end of 7 days of withholding water from
plants droughted at tillering (D1) and equivalent time for well watered (WW) plants 55
Figure 3.6B Water potential (left), osmotic potential (mid) and pressure potential (right) in cv.
Janz (J) infected with <i>Pythium</i> (P), <i>Rhizoctonia</i> (R) and control (C). The measurements
were taken at the beginning and end of 7 days of withholding water from plants
droughted at tillering (D1) and equivalent time for well watered (WW) plants
Figure 3.6C Relative water content (%) for WW (0) and D1 (7) after 7 days of withholding
water for cvs. Mulgara (left) and Janz (right) infected with <i>Pythium</i> (P), <i>Rhizoctonia</i> (R)
and control (C). The measurements were taken at the beginning and end of 7 days of

withholding water from plants droughted at tillering (D1) and equivalent time for well watered (WW) plants
Figure 3.7A Water potential (left), osmotic potential (mid) and pressure potential (right) in cv
Mulgara (M) infected with <i>Pythium</i> (P), <i>Rhizoctonia</i> (R) and control (C). The
measurements were taken at the beginning and end of 7 days of withholding water from
plants droughted at anthesis (D2) and equivalent time for well watered (WW) plants 57
Figure 3.7B Water potential (left), osmotic potential (mid) and pressure potential (right) in cv
Janz (J) infected with <i>Pythium</i> (P), <i>Rhizoctonia</i> (R) and control (C). The measurements
were taken at the beginning and end of 7 days of withholding water from plants
droughted at anthesis (D2) and equivalent time for well watered (WW) plants 57
Figure 3.7C Relative water content (%) for WW (0) and D2 (7) after 7 days of withholding
water for cvs. Mulgara (left) and Janz (right) infected with Pythium (P), Rhizoctonia (R
and control (C). The measurements were taken at the beginning and end of 7 days of
withholding water from plants droughted at anthesis (D2) and equivalent time for well
watered (WW) plants
Figure 3.8A Water use efficiency based on grain yield (g/l) for cvs. Mulgara and Janz infected
by Pythium at either tillering (D1) or anthesis (D2) droughts compared with well
watered (WW) plants. 61
Figure 3.8B Water use efficiency based on grain yield (g/l) for cvs. Mulgara and Janz infected
by <i>Rhizoctonia</i> at either tillering (D1) or anthesis (D2) droughts compared with wel
watered (WW) plants. 61
Figure 3.8C Water use efficiency based on grain yield (g/l) for uninfected cvs. Mulgara and
Janz at either tillering (D1) or anthesis (D2) droughts compared with well watered
(WW) plants
infected by <i>Pythium</i> at either tillering (D1) or anthesis (D2) droughts compared with
well watered (WW) plants
Figure 3.9B Water use efficiency based on shoot dry weight (g/l) for cvs. Mulgara and Janz
infected by <i>Rhizoctonia</i> at either tillering (D1) or anthesis (D2) droughts compared with
well watered (WW) plants.
Figure 3.9C Water use efficiency based on shoot dry weight (g/l) for cvs. Mulgara and Janz
controls at either tillering (D1) or anthesis (D2) droughts compared with well watered
(WW) plants64
Figure 3.10A Effect of diseases (Pythium and Rhizoctonia) on lesion % of Mulgara and Janz
The diseases were compared with controls
Figure 3.10B Effect of droughts on lesion % of Mulgara and Janz. Root lesions for 1st drough
(D1) at tillering and 2 nd drought (D2) at anthesis were compared with well watered
(WW) plants65
Figure 3.11A Roots of Janz (left) and Mulgara (right) at anthesis drought (D2). Roots were
infected by Pythium (right side, Rhizoctonia (mid) and control (left)

Figure 3.11B Roots of Janz (left) and Mulgara (right) at tillering drought (D1). Roots were
infected by Pythium (right side, Rhizoctonia (mid) and control (left)67
Figure 3.11C Roots of Janz (left) and Mulgara (right) at well water (WW). Roots were
infected by Pythium (right side, Rhizoctonia (mid) and control (left)
Figure 4.1 Effect of inoculum density of <i>Pythium</i> on cumulative transpiration for wheat cv.
Janz from tillering (11 January) until harvest (15 March). Values are means of 6
replicates80
Figure 4.2 Effect of inoculum density of <i>Pythium</i> on weekly transpiration rate of wheat cv.
Janz from tillering (11 January) until harvest (15 March)
Figure 4.3 Effect of inoculum density of <i>Pythium</i> on WUE _{grain} and WUE _{DM}
Figure 4.4 Effect of 10 g/pot of <i>Pythium</i> on cumulative transpiration of wheat cv. Janz from
three-leaf stage (17 May) until harvest (14 August). Anthesis started on 25 June. Water
was withheld at late anthesis from 7 July 2010. Values are means of 12 replicates 85
Figure 4.5 Effect of a 10 g/pot inoculum density of Pythium on transpiration per week of
wheat cv. Janz from the three-leaf stage (17 May) until harvest (14 August). Values are
means of 12 replicates
Figure 4.6 Effect of 10g/ pot of Pythium on water use efficiency of cv. Janz based on both
grain yield (WUE grain) and shoot dry matter (WUE DM). Values are means ± s.e., n=12.
Figure 4.7 Predawn water potential (Ψ) for wheat cv. Janz for both controls and <i>Pythium</i> at 0,
7, 14 and 21 days after withholding water. Values are means \pm s.e., n=3
Figure 4.8 Midday water potential (MPa) for cv. Janz for both controls and <i>Pythium</i> at 0, 7, 14
and 21 days after withholding water. Values are means \pm s.e., n=390
Figure 4.9 Relationships between predawn and midday water potentials for (a) uninoculated
and (b) inoculated plants during 21 days of withholding water
Figure 4.10 Photosynthetic rate (μmol CO ₂ m ⁻² s ⁻¹) of controls and <i>Pythium</i> of cv. Janz after 0,
7, 14 and 21 days of withholding water. Values are means \pm s.e., n=6
Figure 4.11 Stomatal conductance (mol H ₂ O m ⁻² s ⁻¹) of controls and <i>Pythium</i> of wheat cv.Janz
after 0, 7, 14 and 21 days of withholding water. Values are means \pm s.e., n = 6 93
Figure 4.12 Intercellular CO ₂ concentration (µmol CO ₂ mol ⁻¹) of controls and <i>Pythium</i> of
cv.Janz after 0, 7, 14 and 21 days of water withholding. Values are means \pm s.e., n =694
Figure 4.13 Transpiration rate (mmol H ₂ O m ⁻² s ⁻¹) of controls and <i>Pythium</i> of cv. Janz after 0,
7, 14 and 21 days of withholding water. Values are mean \pm s.e., n =6
Figure 4.14 Instantaneous water use efficiency (mmol CO ₂ /mol H ₂ O) of controls and
Pythium of cv. Janz after 0, 7, 14 and 21 days of withholding water. Values are mean \pm
s.e., n = 6
Figure 4.15 The relationship between photosynthesis (μ mol CO ₂ m ⁻² s ⁻¹) and transpiration rate
(mmol H ₂ O m ⁻² s ⁻¹) as affected by <i>Pythium</i> inoculation for wheat cv. Janz during the 21
day water withholding period after anthesis. R^2 are significant at $P < 0.01$

Figure 4.16 The relationship between photosynthesis (μ mol CO ₂ m ⁻² s ⁻¹) and stomatal conductance (mol H ₂ O m ⁻² s ⁻¹) as affected by <i>Pythium</i> inoculation for cv. Janz during the 21 day water withholding period after anthesis. R ² are significant at $P < 0.01 \dots 97$
Figure 4.17 Stomatal conductance (mol H ₂ O m ⁻² s ⁻¹) for uninoculated and inoculated plants in relation to midday leaf water potential (MPa) during the 21 day water withholding period after anthesis.
Figure 4.18 The relationship between water use efficiency $intrinsic$ (A/g _s) and stomatal
conductance (g _s) for uninoculated and inoculated plants during the 21 day water withholding period after anthesis
Figure 4.19 The relationship between water use efficiency <i>instantaneous</i> (A/E) and stomatal conductance (g _s) for uninoculated and inoculated plants during the 21 day water
withholding period after anthesis
Figure 5.1 Growth of <i>Pythium</i> on roots and root lesion (arrows) of wheat inoculated by
Pythium in hydroponic solution
Figure 5.2 Root pruning (Rp) and <i>Pythium</i> (<i>P</i>) effects on transpiration per day of cv. Janz at 4
days before inoculation and 10 days after inoculation in absence of Polyethyleneglycol
(-PEG) and presence of PEG-induced drought (+PEG) (1 st experiment). Arrow shows
when PEG and root pruning treatments started
Figure 5.3 Root pruning (Rp) and <i>Pythium</i> (P) effects on transpiration per day of cv. Janz at 5
days before inoculation and 14 days after inoculation in absence of Polyethyleneglycol
(-PEG) and presence of PEG-induced drought (+PEG) (2 nd experiment). Arrow shows
when PEG and root pruning treatments started
Figure 5.4 Comparison of transpiration per week (TPW) between all treatments on 7 and 14
August (1 st experiment). Rp= root pruning, P=Pythium and D=PEG 121
Figure 5.5 Comparison of transpiration per week (TPW) between all treatments on 2 and 9
October (2 nd experiment). Rp= root pruning, P=Pythium and D=PEG
Figure 5.6 Root pruning and <i>Pythium</i> effects on (A) WUE (shoot + root) and (B) WUE (shoot) of cv.
Janz in absence of Polyethyleneglycol (-PEG) and presence of PEG-induced drought
(+PEG) in first experiment. WUE was measured in g/l. Each column represents the
mean \pm s.e for eight plants. 125
Figure 5.7 Root pruning and <i>Pythium</i> effects on (A) WUE (shoot + root) and (B) WUE (shoot) of cv.
Janz in absence of polyethyleneglycol (-PEG) and presence of PEG-induced drought
(+PEG) in second experiment. Each data represents the mean ±s.e for eight plants 126
Figure 5.8 Root pruning and <i>Pythium</i> effects on (A) Predawn and (B) Midday water potential
of cv. Janz in absence of polyethyleneglycol (-PEG) and presence of PEG-induced
drought (+PEG). Each column represents the mean ±s.e for four plants
(C) Relative water content of cv. Janz in absence of polyethyleneglycol (-PEG) and
presence of PEG-induced drought (+PEG). Each column represents the mean ±s.e for
four plants